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# Biological effects of ultra-low radiation exposure

Chris Thome  
Dr. Douglas Boreham

SNOLAB Future Projects Workshop  
August 25, 2015

# Research group



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## Dr. Douglas Boreham

Professor and Division Head of Medical Sciences – NOSM  
Adjunct Professor – McMaster University  
Principal Scientist – Bruce Power

### Research interests:

- Low-dose radiobiology
- Diagnostic imaging
- Cancer therapy

## Chris Thome

Post doctoral researcher - NOSM

## Jake Pirkkanen

Graduate student – Laurentian University

## Andrew Zarnke

Graduate student – Laurentian University

# Funding



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## Bruce Power industrial support

- 2015: \$85,000
- 2016-2020: \$1,000,000 (\$200,000 per year)

## NSERC discovery

- 2015-2020: \$190,000 (\$38,000 per year)

## Mitacs Accelerate

- 2015-2017: \$330,000

## NSERC CRD

- 2016-2020: \$1,000,000 (\$200,000 per year)

# Rationale



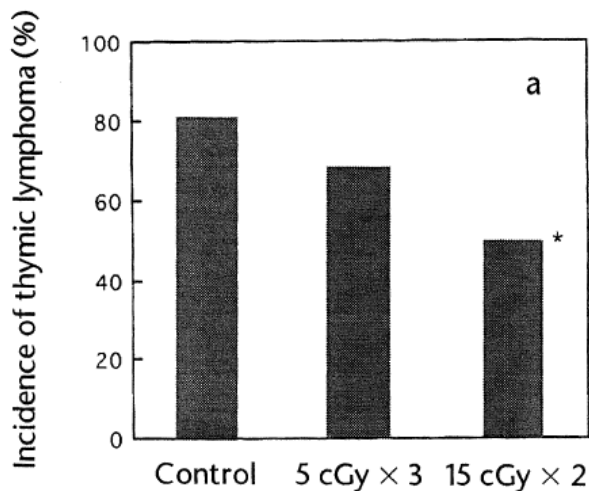
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- Ionizing radiation is ubiquitous and all living organisms on earth
- There is increasing concern over radiation exposure from medical diagnostic procedures
- The effects from these exposures still remains largely unknown
- Limited epidemiological data exists in the low-dose region ( $< 100$  mGy)
- There is growing evidence to suggest that low-dose radiation may provide beneficial effects to living systems
- SNOLAB provides a unique environment to examine the effects of ultra-low background radiation exposure

# Low-dose and cancer

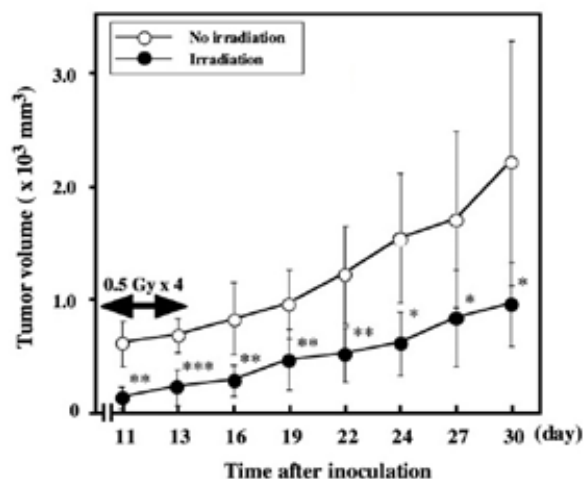


## Reduction in cancer incidence



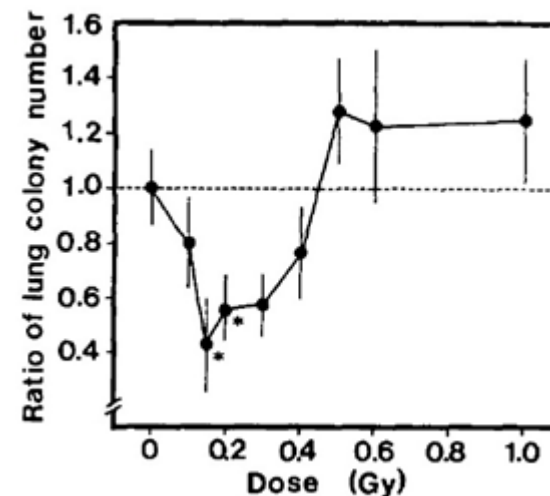
Ishii *et al* 1996

## Decrease in primary tumor growth



Kojima *et al* 2004

## Reduction in tumor metastases



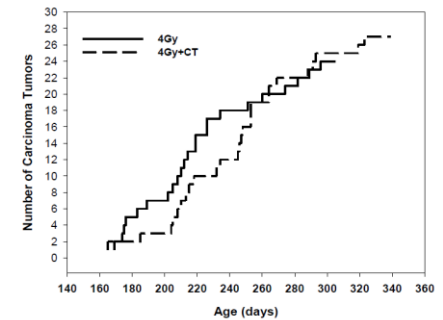
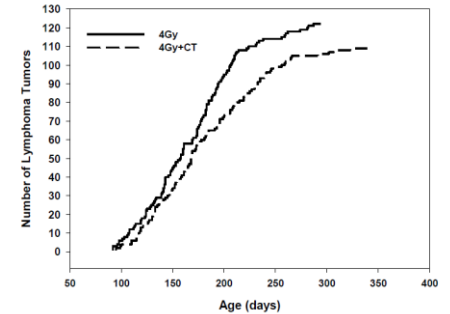
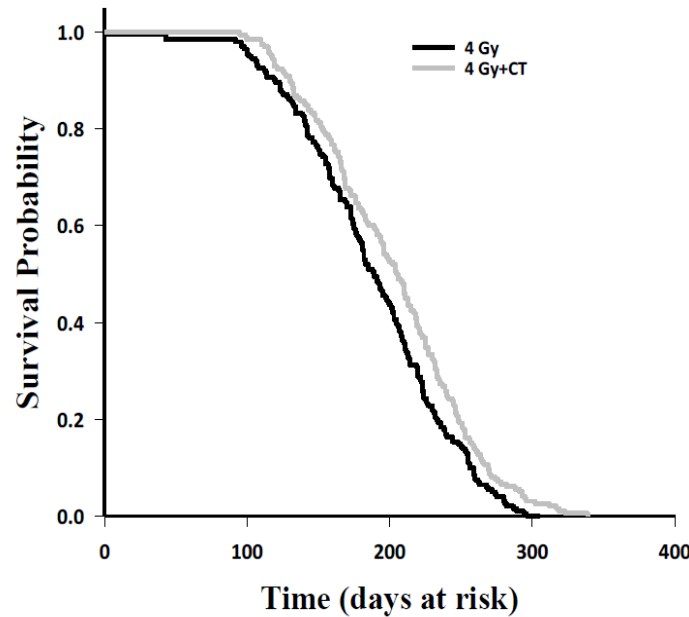
Hosoi and Sakamoto 1993

# Diagnostic imaging



## CT Scan

- Increased mean survival time
- Increased latency of lymphomas and carcinomas



Phan *et al*

## PET Scan

- Reduction in kidney disease

Table IV. Number of *Trp53*<sup>+/-</sup> mice with tissue-specific lesions

Treatment	Kidneys	Bladder	Heart	Reproductive organs
Control	32	3	0	28
10 mGy $\gamma$ -rays	23	6	0	20
10 mGy PET	15*	0	0	27
4 Gy $\gamma$ -rays	25	0	0	38
10 mGy PET + 4 Gy $\gamma$ -rays	32	0	0	36

\* $P < 0.021$  relative to unirradiated control mice.

Taylor *et al* 2014

# Growth stimulation

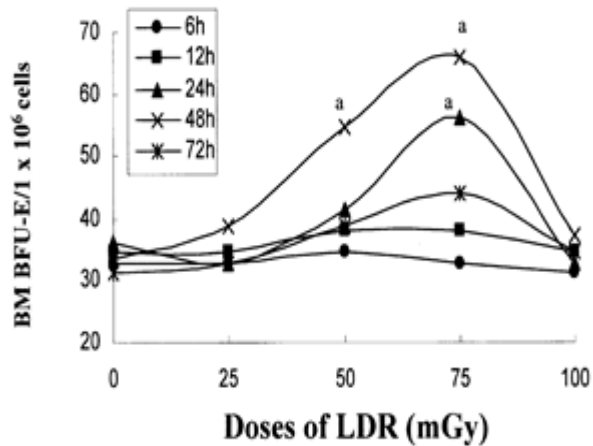


Increased body size in amphibians irradiated during embryogenesis

(Stark *et al* 2015)

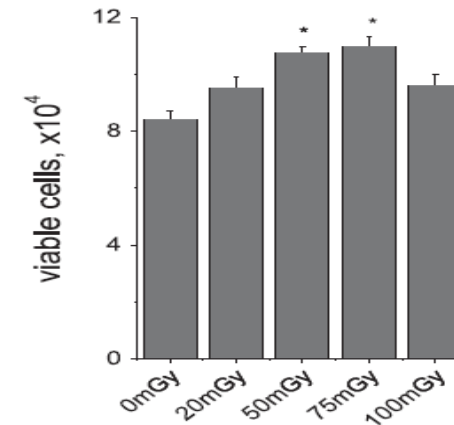
Increased plant size when seeds irradiated (Miller and Miller 1987)

Bone marrow hematopoietic progenitor cells



Li *et al* 2004

Cultured mesenchymal stem cells



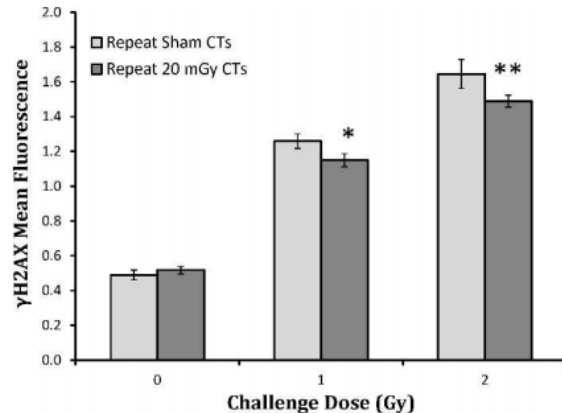
Liang *et al* 2011

# Adaptive response



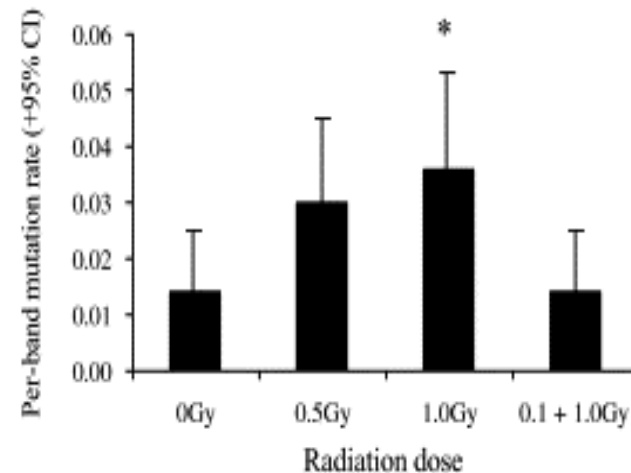
Low-dose radiation exposure can protect against a future high-dose exposure

## DNA DSB formation



Phan *et al* 2012

## Mutation rate



Somers *et al* 2004



# Hypothesis and Objectives



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## Hypothesis:

Natural background radiation is essential for life and maintains genomic stability in living organisms

Prolonged exposure to ultra-low radiation environments will be detrimental to biological systems

## Objectives:

1. Establish a functional biological research laboratory within SNOLAB
2. Examine the effects of incubation in SNOLAB compared to surface control laboratory using 2 simple model systems
  - Cell culture – C3H 10T1/2 cell line
  - Whole organism – Lake Whitefish embryonic development

# Low background results



1. Removal of natural background radiation impairs growth. Growth rates are restored once radiation is artificially reintroduced. Demonstrated with:

- Paramecium shielded with lead (Planel *et al* 1976)
- Blue-green algae (*Synechococcus lividus*) shielded with lead (Conter *et al* 1983)
- Yeast (*Saccharomyces cerevisiae*) shielded with lead/cadmium (Gajendiran and Jeevanram 2002)
- Bacteria (*Deinococcus radiodurans*) grown in Waste Isolation Pilot Plant (WIPP) (Smith *et al* 2011)
- Mouse lymphoma L5178Y cells shielded with lead (Taizawa *et al* 1992)
- Mouse lymphoma L5178Y cells shielded with iron (Kawanishi *et al* 2012)

# Low background results



## 2. Removal of natural background radiation reduces repair capacity towards induced damage. Demonstrated with:

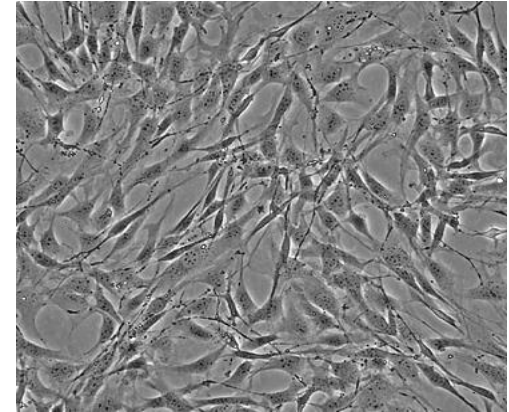
- Survival fraction in yeast (*Saccharomyces cerevisiae*) shielded with lead/cadmium (Gajendiran and Jeevanram 2002)
- Background/induced mutation rate in Chinese hamster V79 cells grown in Gran Sasso Underground Laboratory (LNGS) (Satta *et al* 2002)
- Micronuclei formation and ROS scavenging in human lymphoblastoid TK6 cells grown in LNGS (Carbone *et al* 2010)

# C3H 10T1/2 cell line



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- Mouse embryonic stem cell line
- Pre-carcinogenic
- High spontaneous transformation rate
- Sensitive to low-dose ionizing radiation



## Previous findings:

A single low-dose of radiation (1 mGy to 100 mGy) can decrease transformation frequency below spontaneous levels (*Azzam et al 1996*)

Chronic adapting low-doses of radiation (0.1 Gy, 0.65 Gy or 1.5 Gy) can protect against an acute challenge dose of 4 Gy (*Azzam et al 1994*)

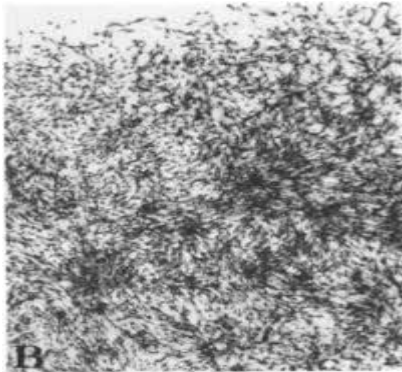
# C3H 10T1/2 cell line



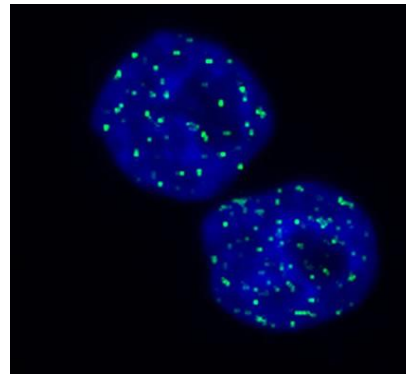
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## Radiobiological endpoints

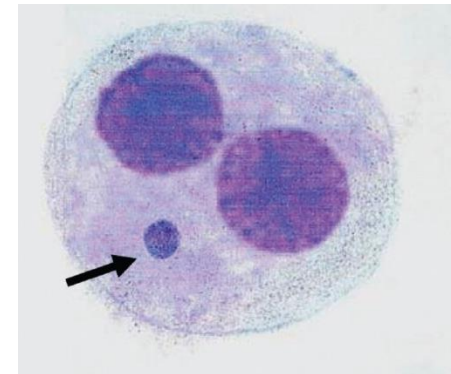
Transformation  
frequency



DNA DSB  
formation



Micronuclei  
formation



# C3H 10T1/2 cell line



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- Cells will be cultured within SNOLAB and the surface control lab
- Glove box incubators enable matching conditions by controlling air, temperature and pressure
- Cells will be cultured for multiple passages and at periodic intervals tested for:
  - Spontaneous transformation frequency
  - Background levels of DNA DSBs and micronuclei
- The dose-response for induced damage will be examined in low-background adapted cells



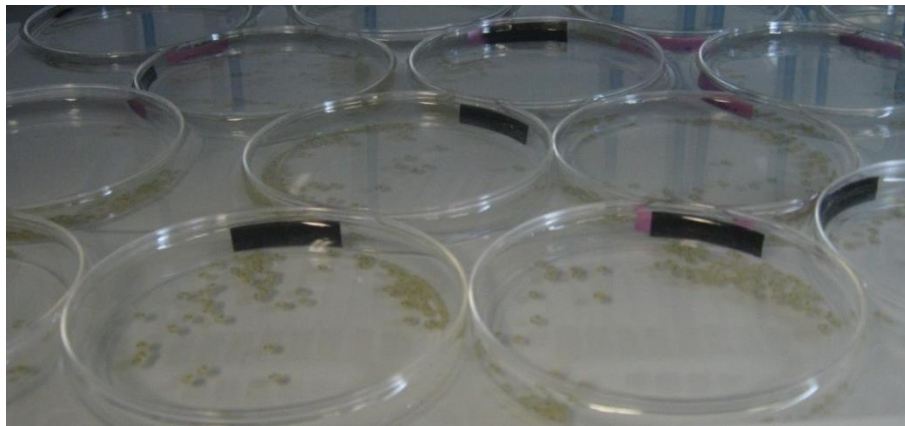
# Lake Whitefish



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Good model organism for examining radiological effects

- Embryogenesis one of the most sensitive life stages to radiation
- Long development period (> 200 days)
  - Extended low-dose chronic exposures
  - Accurate targeting of specific development stages
- Can accurately quantify growth efficiency
- Easy to raise and low maintenance

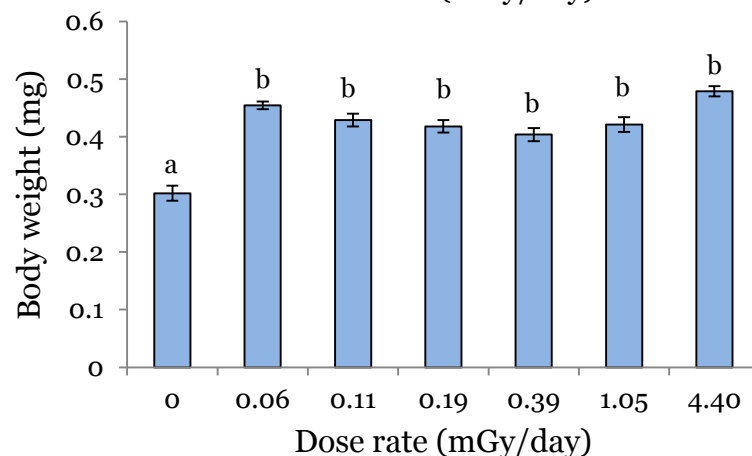
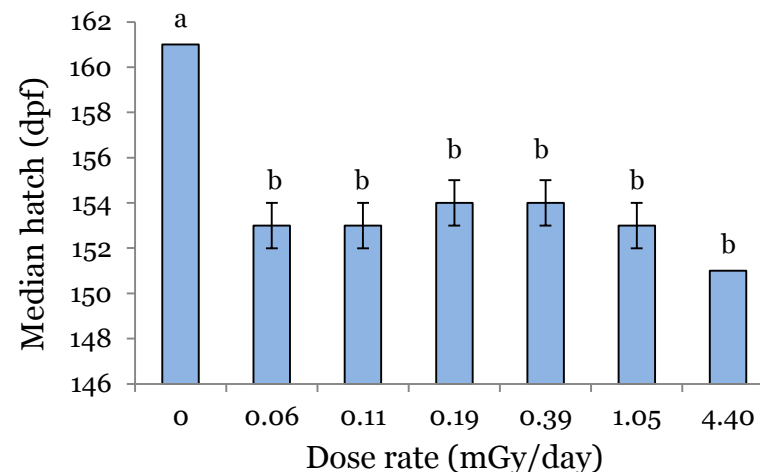
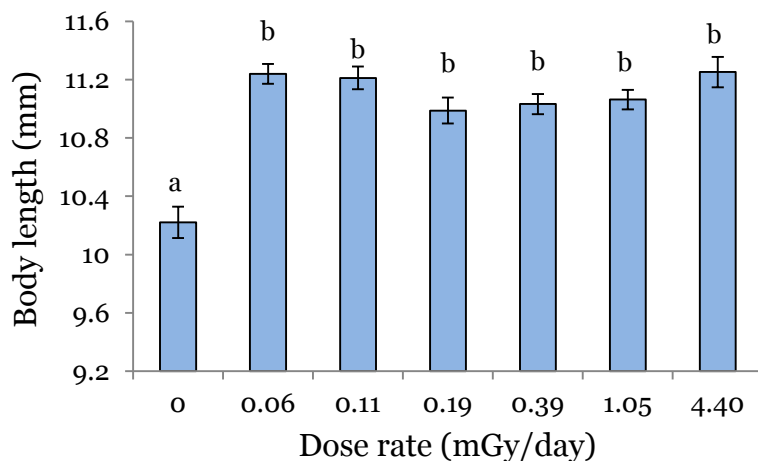


# Lake Whitefish



Chronic low dose  $^{137}\text{Cs}$  gamma ray exposure

- Accelerated development – earlier time to median hatch
- Stimulated growth – larger body length and weight





# Lake Whitefish



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- Lake Whitefish embryos will be reared within SNOLAB and the surface control lab
- Embryos will be raised from fertilization to hatch within standard refrigeration units
- At multiple stages, embryos will be analyzed for
  - Mortality
  - Development rate
  - Size
  - Growth efficiency
- The response to acute thermal or chemical stress will be examined in low-background adapted embryos

