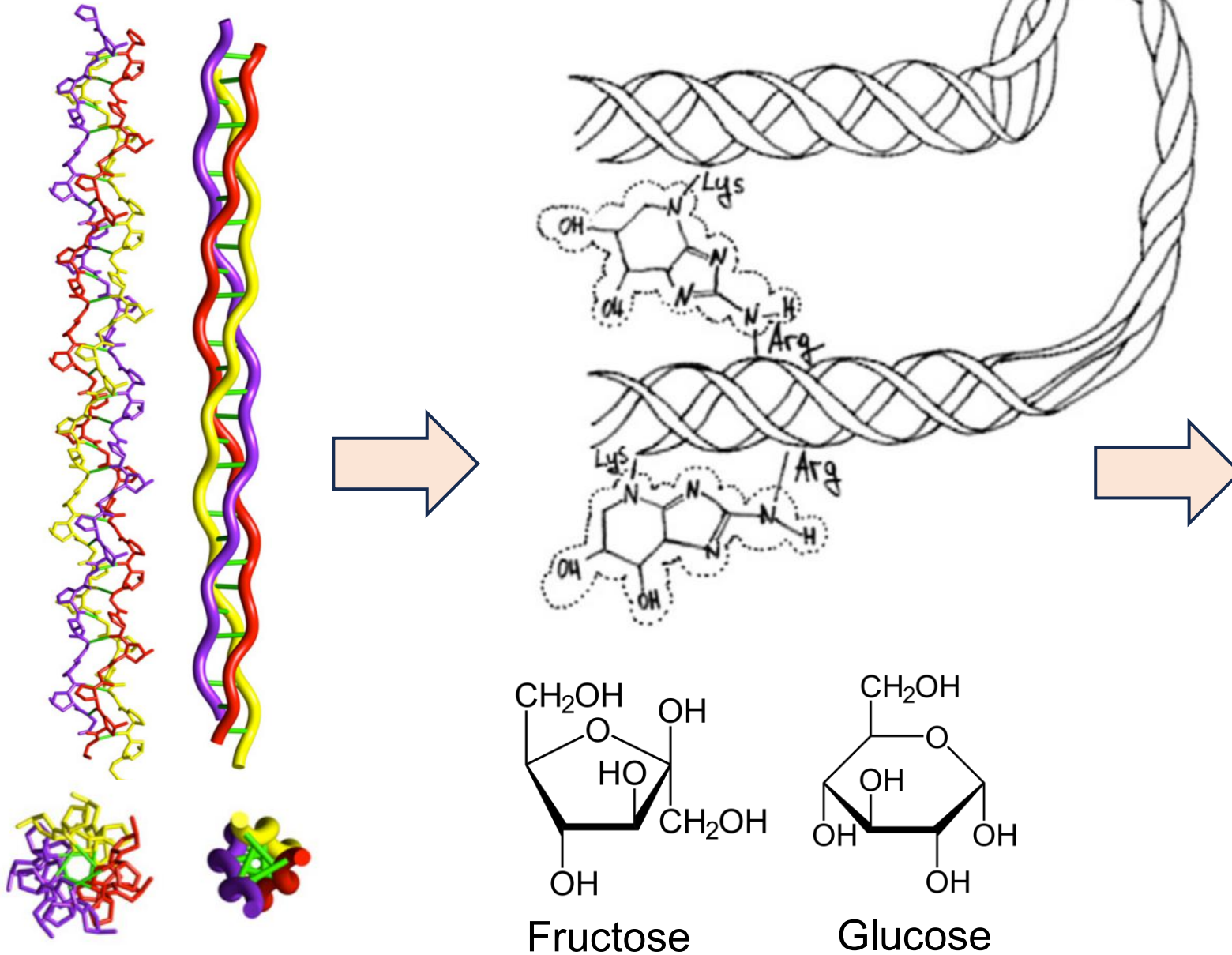


# **Polarization Resolved Second Harmonic Generation Microscopy Reveals Molecular Disorder in Crosslinked Collagen Fibrils**

**Benjamin Hansson**  
Richard Cisek  
Danielle Tokarz  
Laurent Kreplak



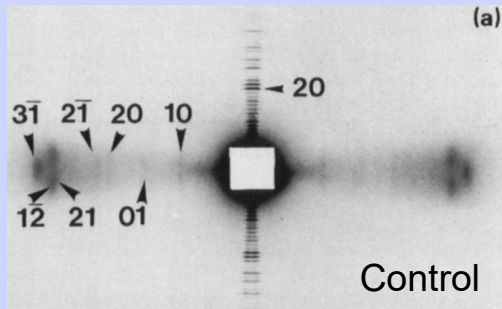
# The AGEing of Collagen



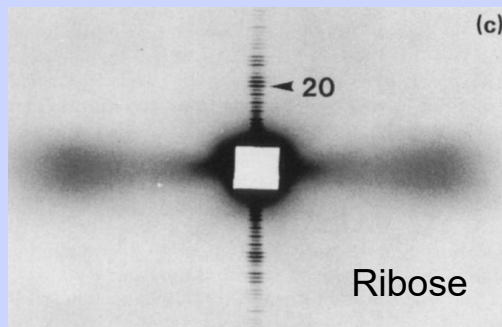
- Age-related diseases
- Cellular dysfunction
- Tendon stiffening
- Impaired wound healing
- Wrinkles

Jordi Bella, Biochemical Journal, (2016)  
A. Gautieri et. al., Matrix Biology, (2014)

# Previous Studies on Collagen Glycation



Control

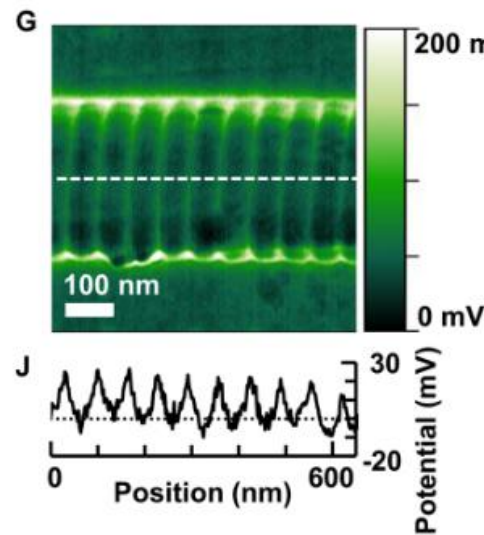


Ribose

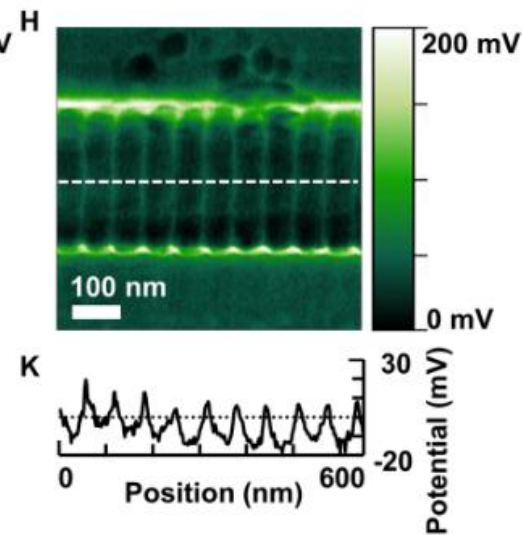
## X-ray diffraction

6 weeks 0.2 M ribose  
(Tanaka et al., 1988)

## Control

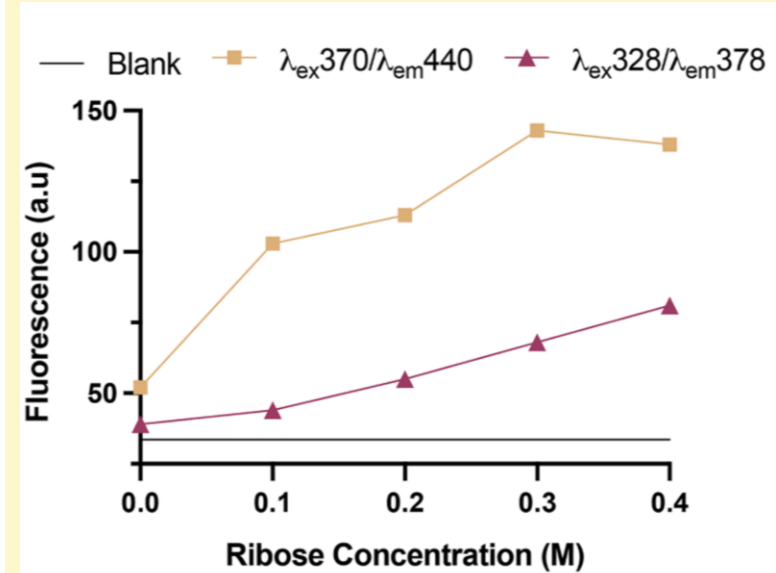


## R5P



## Atomic force microscopy

5 weeks 50 mM  
ribose-5-phosphate  
(Bansode et al., 2020)



## Fluorescence

## spectroscopy

50 weeks ribose  
(Sloseris et al., 2025)

*How can we rapidly  
measure **nanoscale  
changes** in collagen  
structure due to glycation-  
induced crosslinking?*

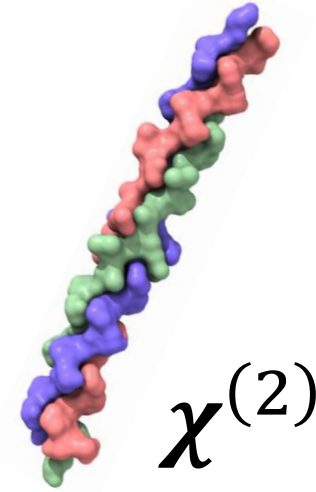
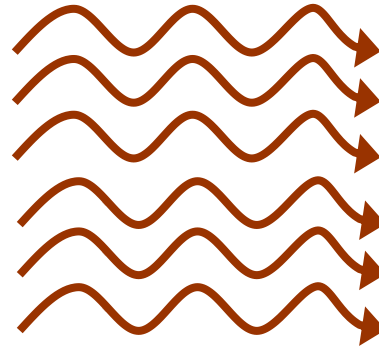
# Second Harmonic Generation (SHG) of Collagen

SHG occurs when two photons convert into one photon of double frequency **only** for media with non-centrosymmetric structure

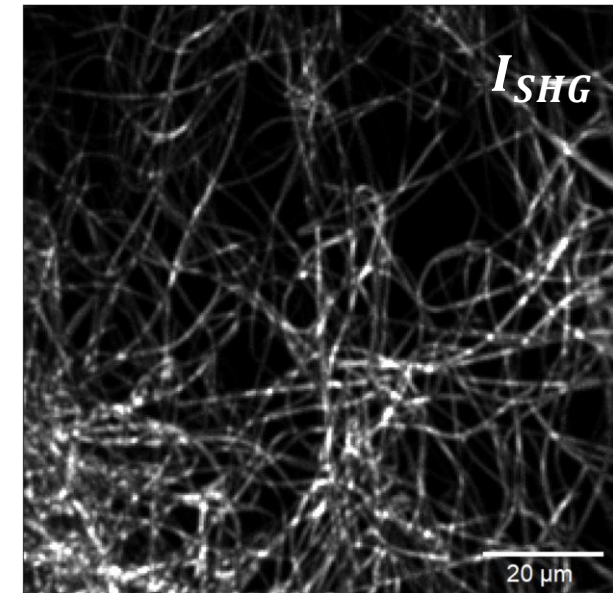
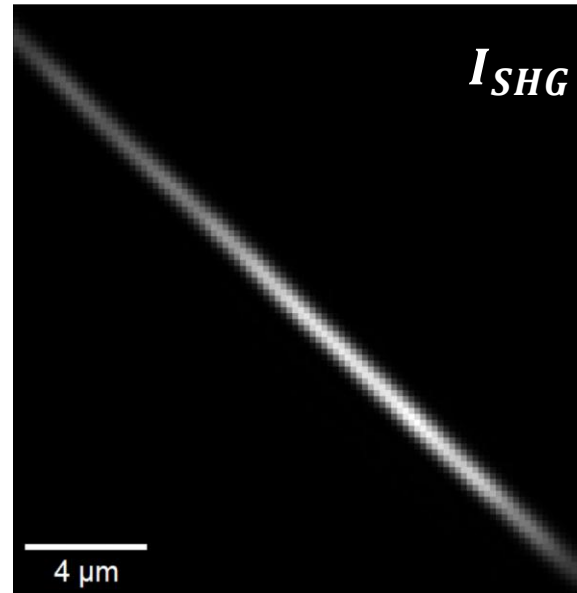
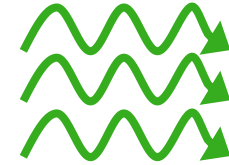
$$I_{SHG} \propto \chi^{(2)} EE$$

$\chi^{(2)}$  is a material property which can be measured by analyzing the polarization of SHG light

1030 nm excitation



515 nm SHG

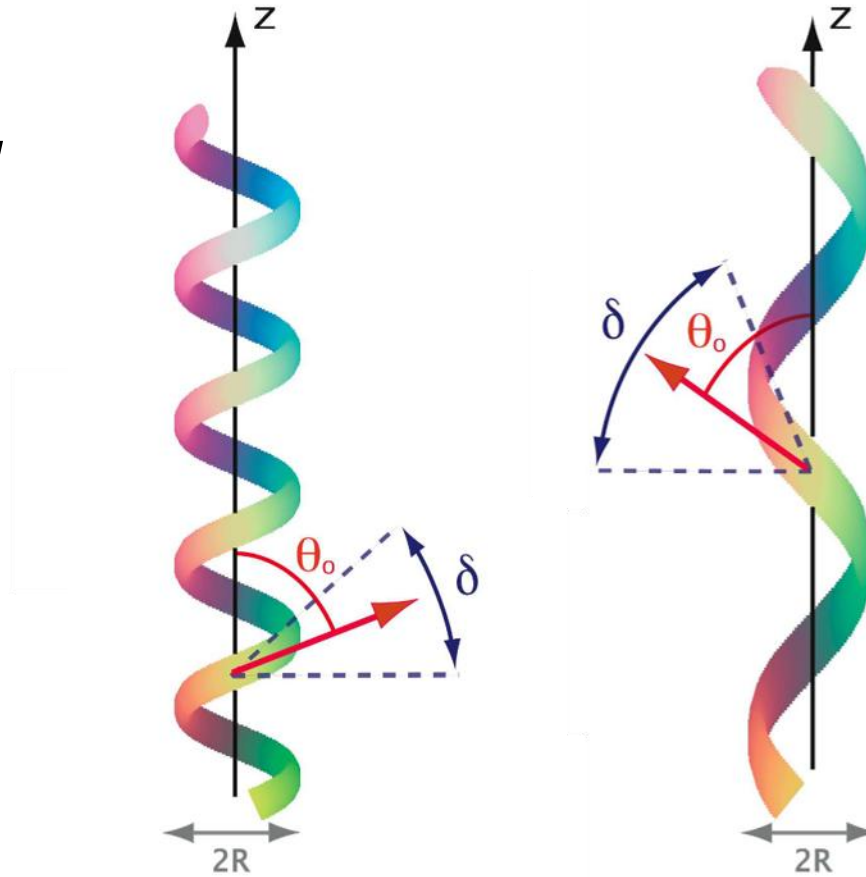


# Using SHG to Investigate Molecular Structure

$$I_{SHG} \propto \chi^{(2)} \mathbf{E} \mathbf{E}$$

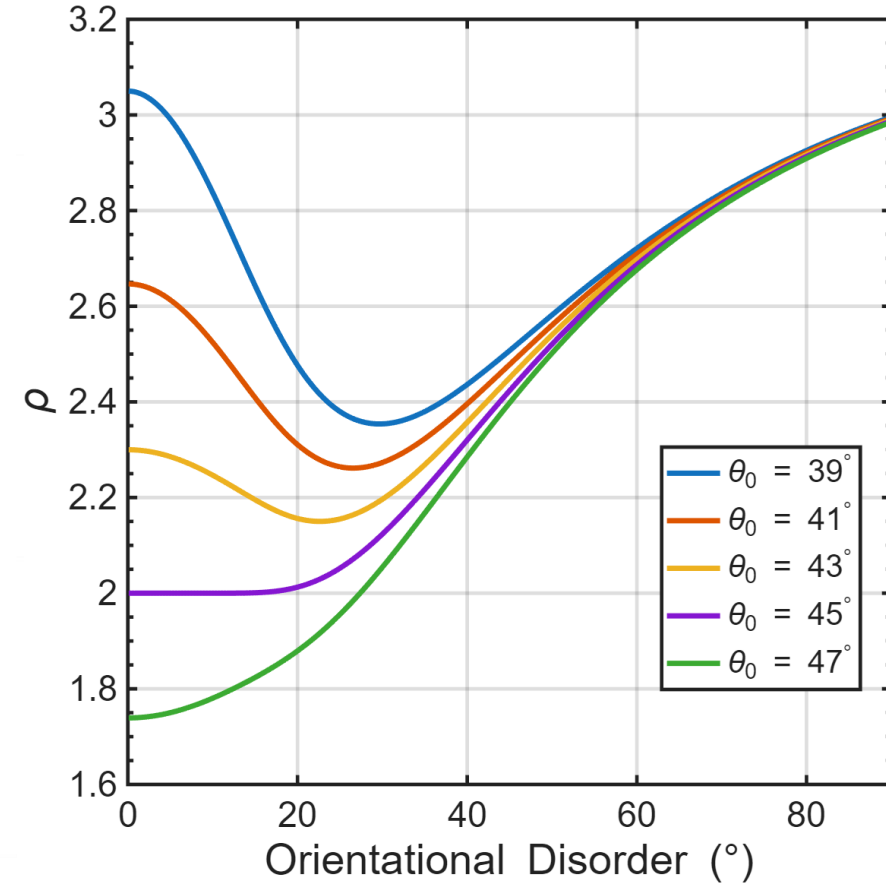
$$\rho = \frac{\chi_{ZZZ}^{(2)}}{\chi_{ZXX}^{(2)}}$$

$$\cos^2 \theta_0 \cong \frac{\rho}{\rho + 2}$$



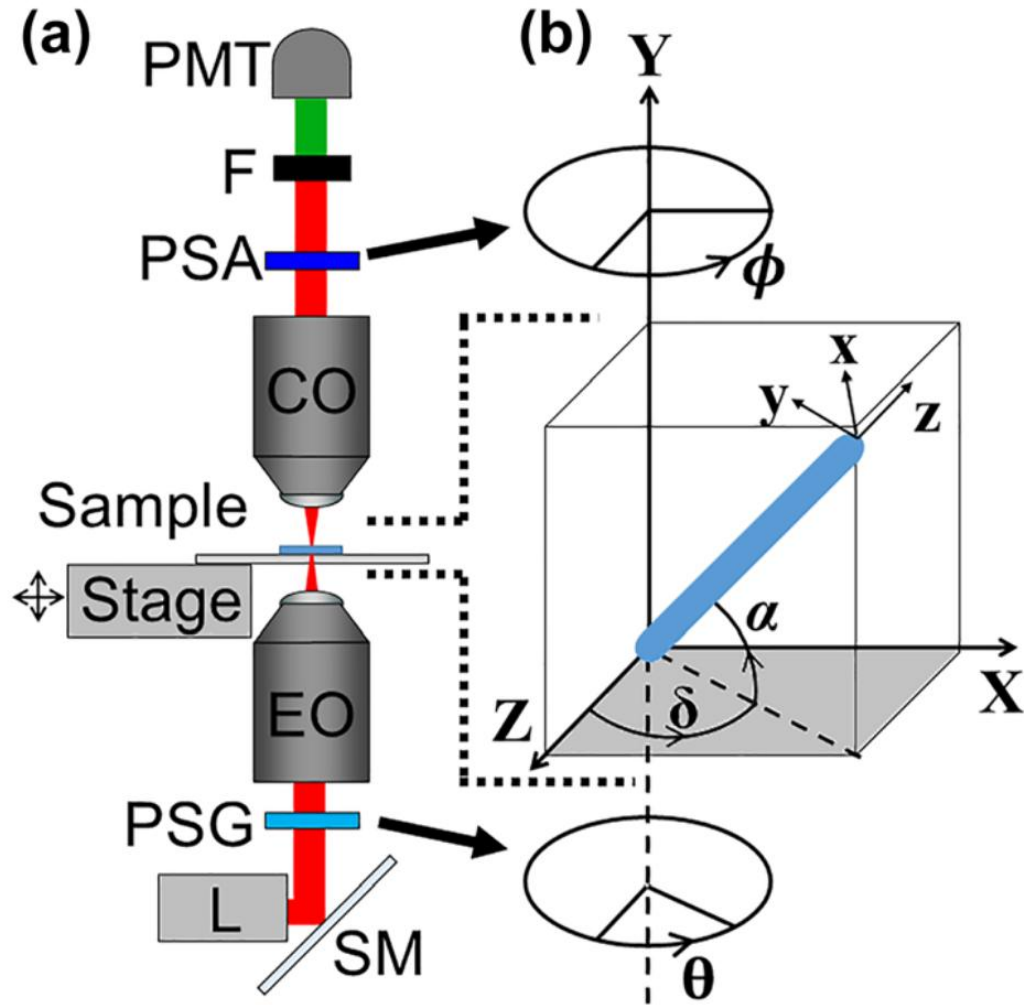
Low  $\rho$   
Compact helix

High  $\rho$   
Stretched helix

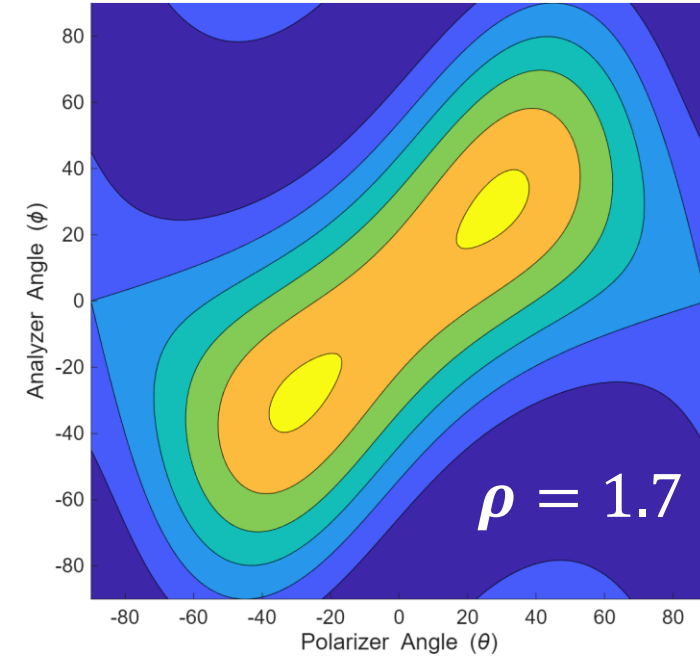
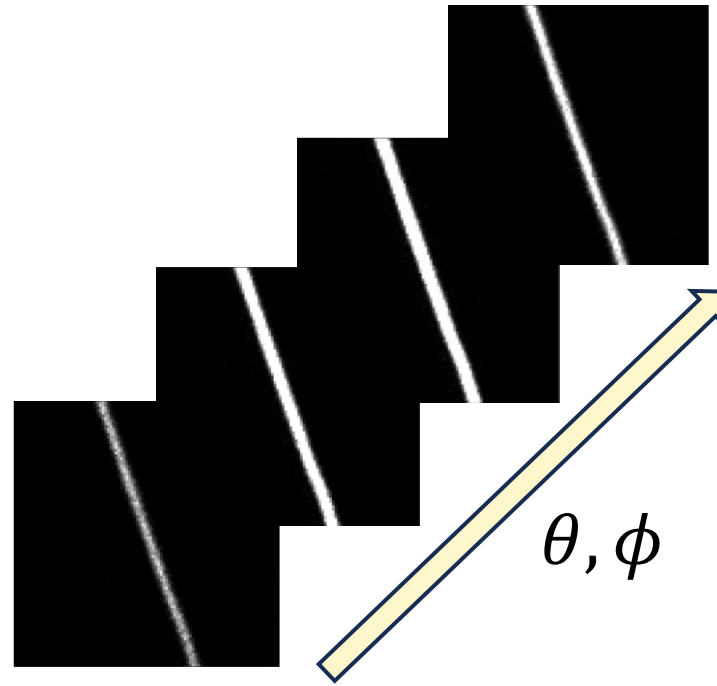


Tiaho et. al., Optics Express, (2007)

# The Polarization-In Polarization-Out (PIPO) SHG Microscope



Harvey et. al., Nanophotonics, (2023)



$$I_{2\omega}^{C6V} \propto |\sin \phi \sin 2\theta - \cos \phi \sin^2 \theta + \rho \sin \phi \cos^2 \theta|^2$$

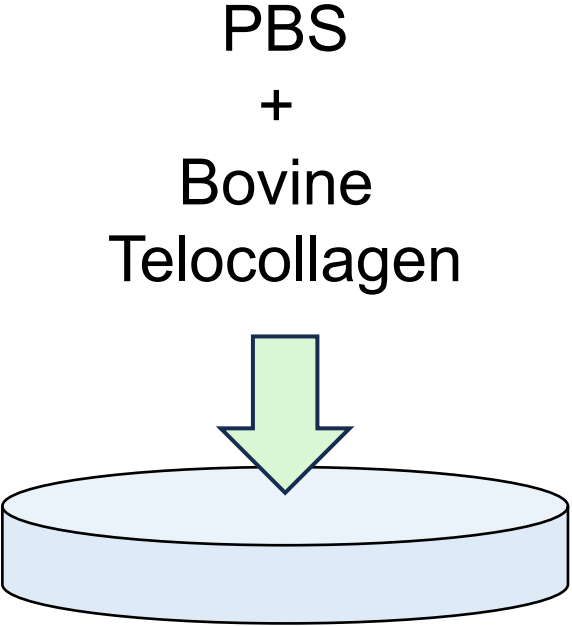
## Research Question 1

*Can **SHG** measure the difference between crosslinked and non-crosslinked collagen?*

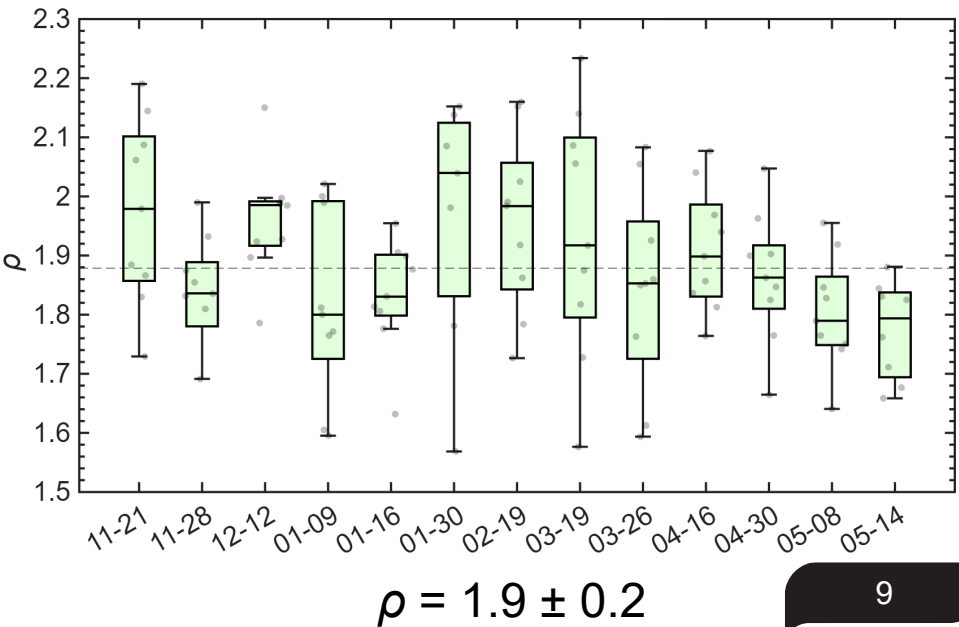
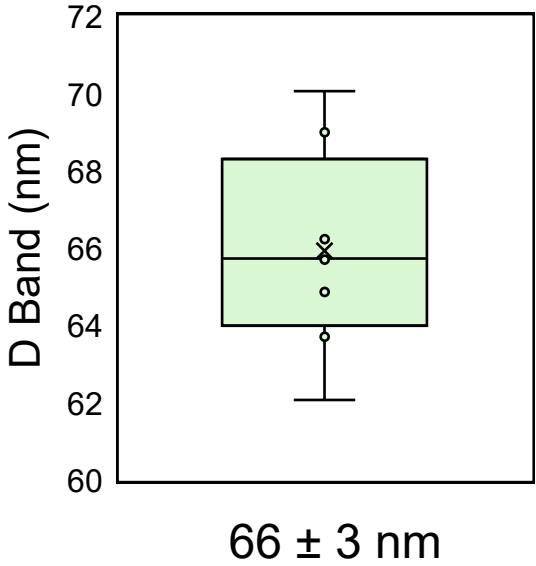
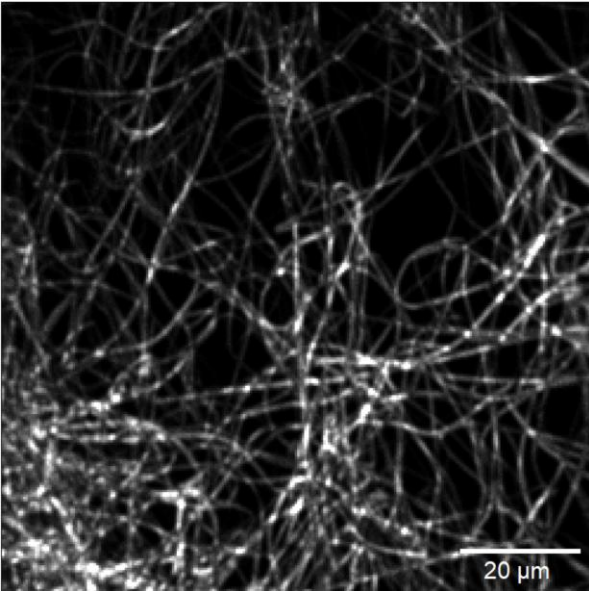
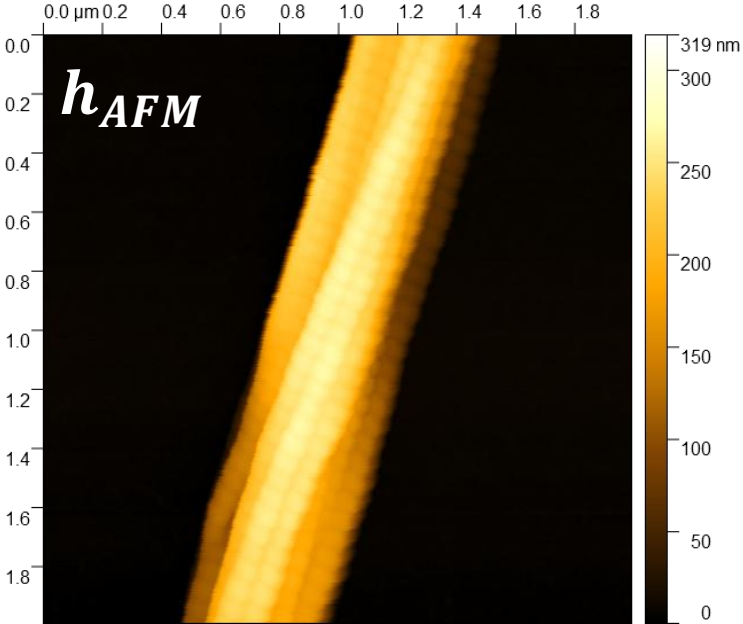
## Research Question 2

*What is the mechanism behind the glycation-induced change in collagen **molecular structure**?*

# A Crosslinking-Free *In-Vitro* Fibril Platform

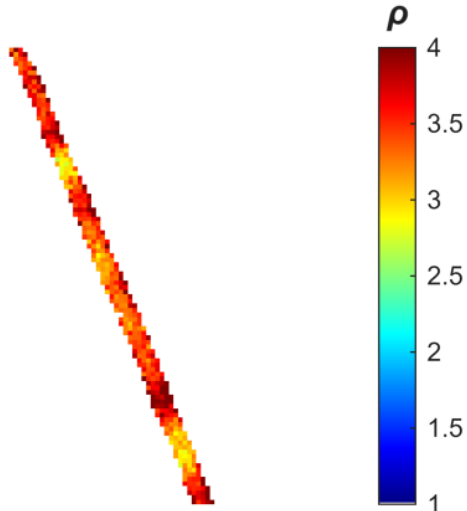


24 hours  
incubation  
at 30°C

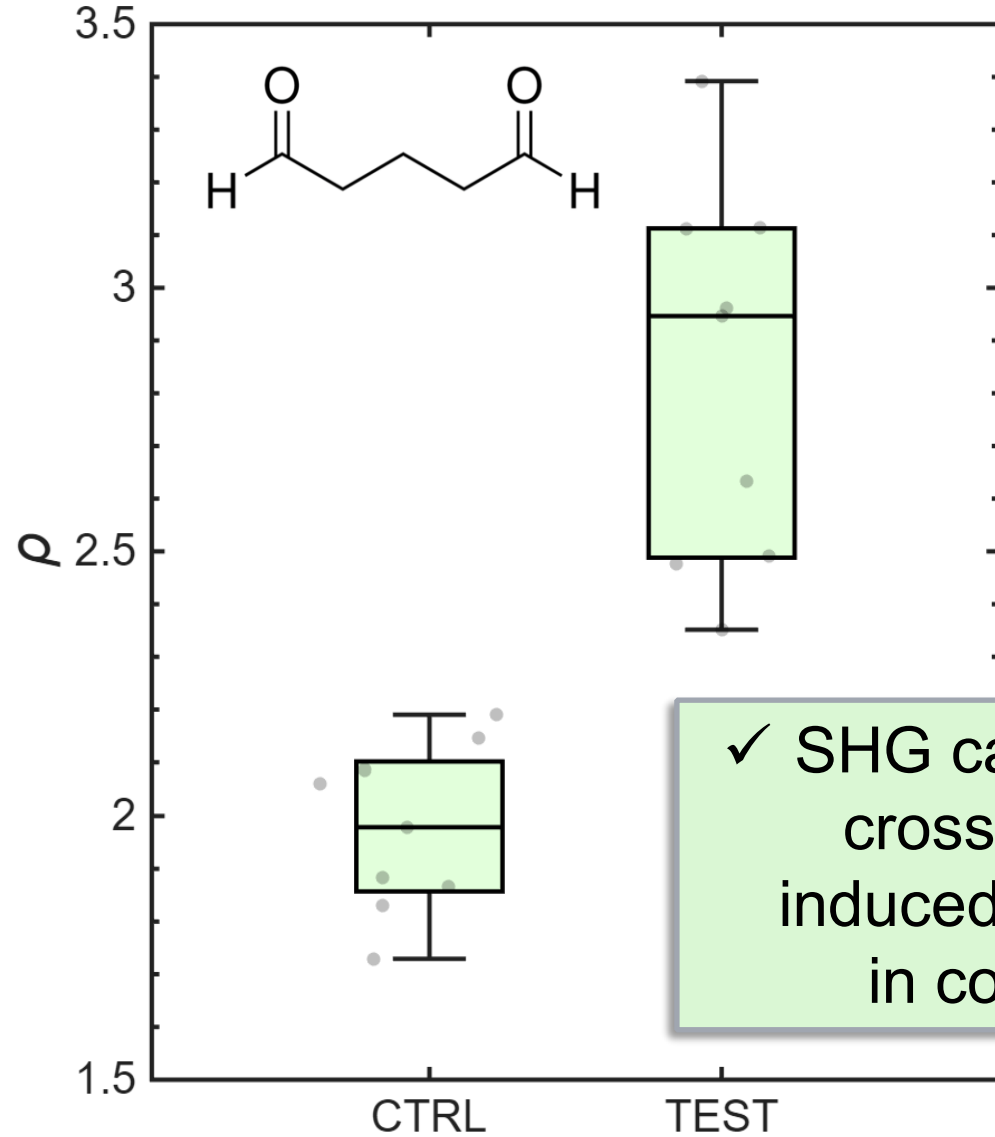
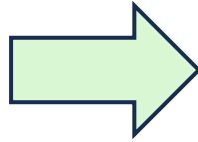
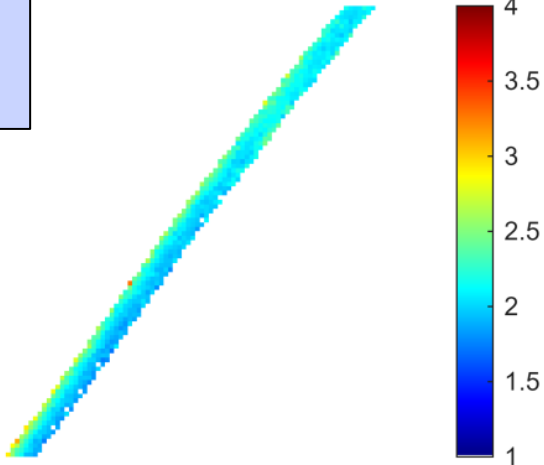


# Using Glutaraldehyde to Induce Crosslinking

Glutaraldehyde  
0.2 M 1 hour  
 $n = 9$



PBS Control  
 $n = 9$



✓ SHG can detect crosslinking-induced changes in collagen

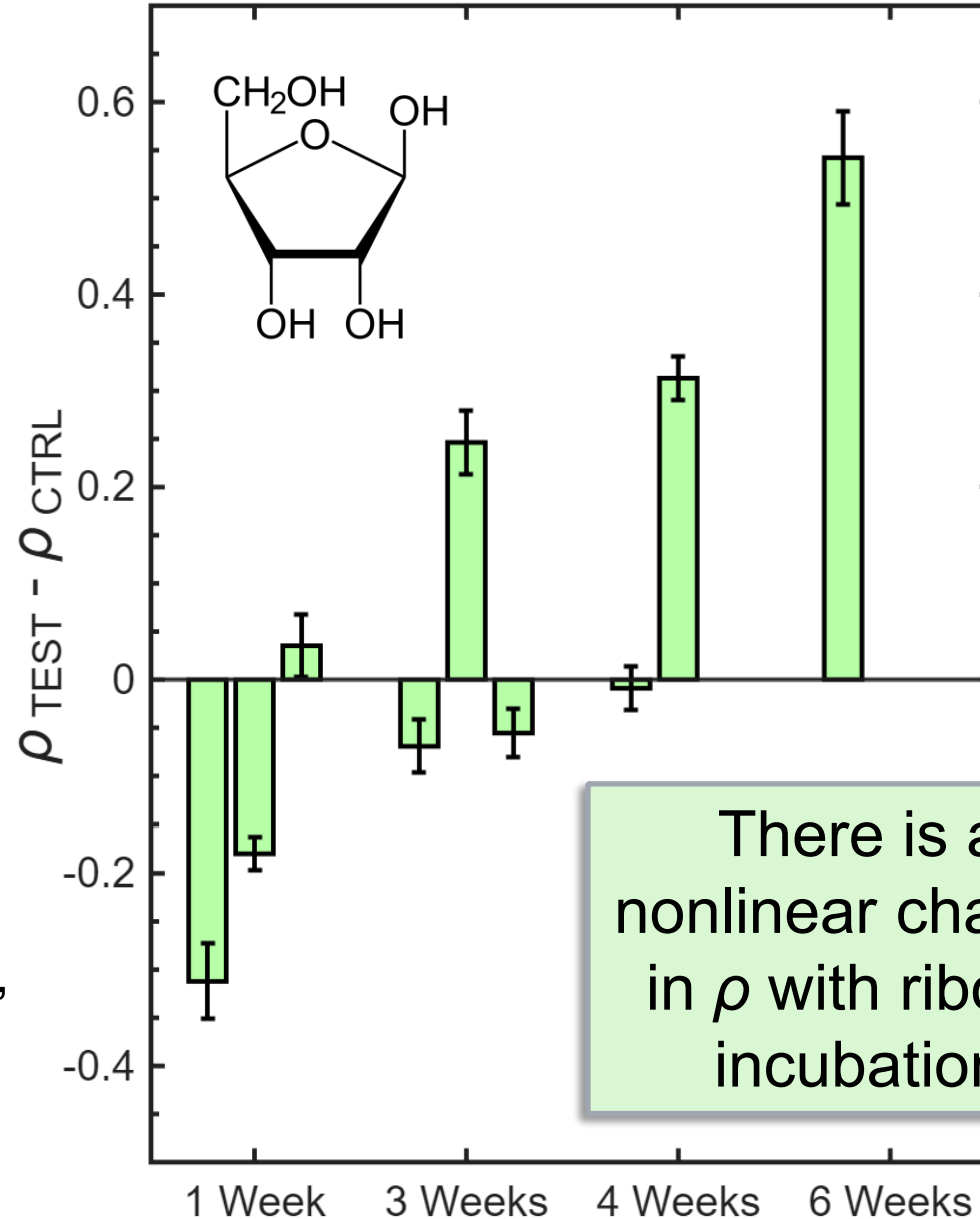
# Nonlinear Change in $\rho$ with Ribose

**Goal:** Measure ribose-treated fibrils at different time points to search for changes in molecular structure

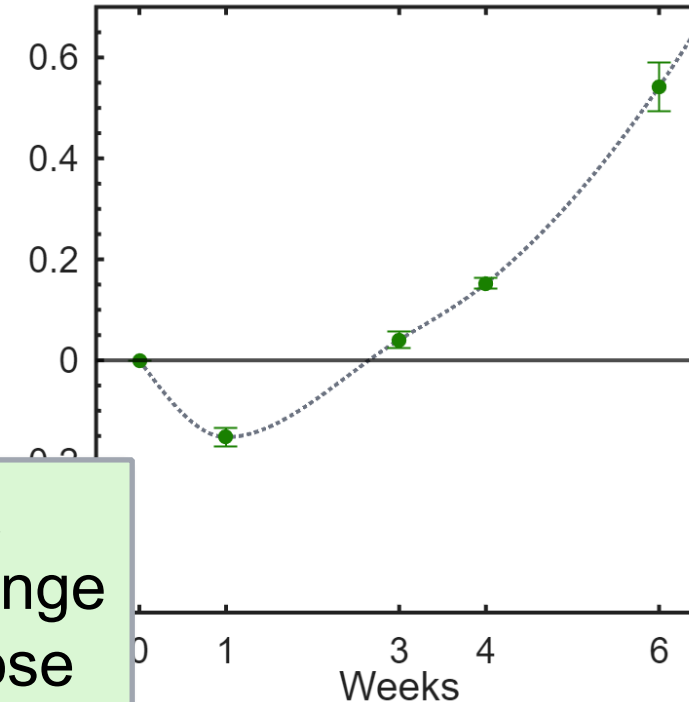
Ribose  
0.2 M

PBS  
Control

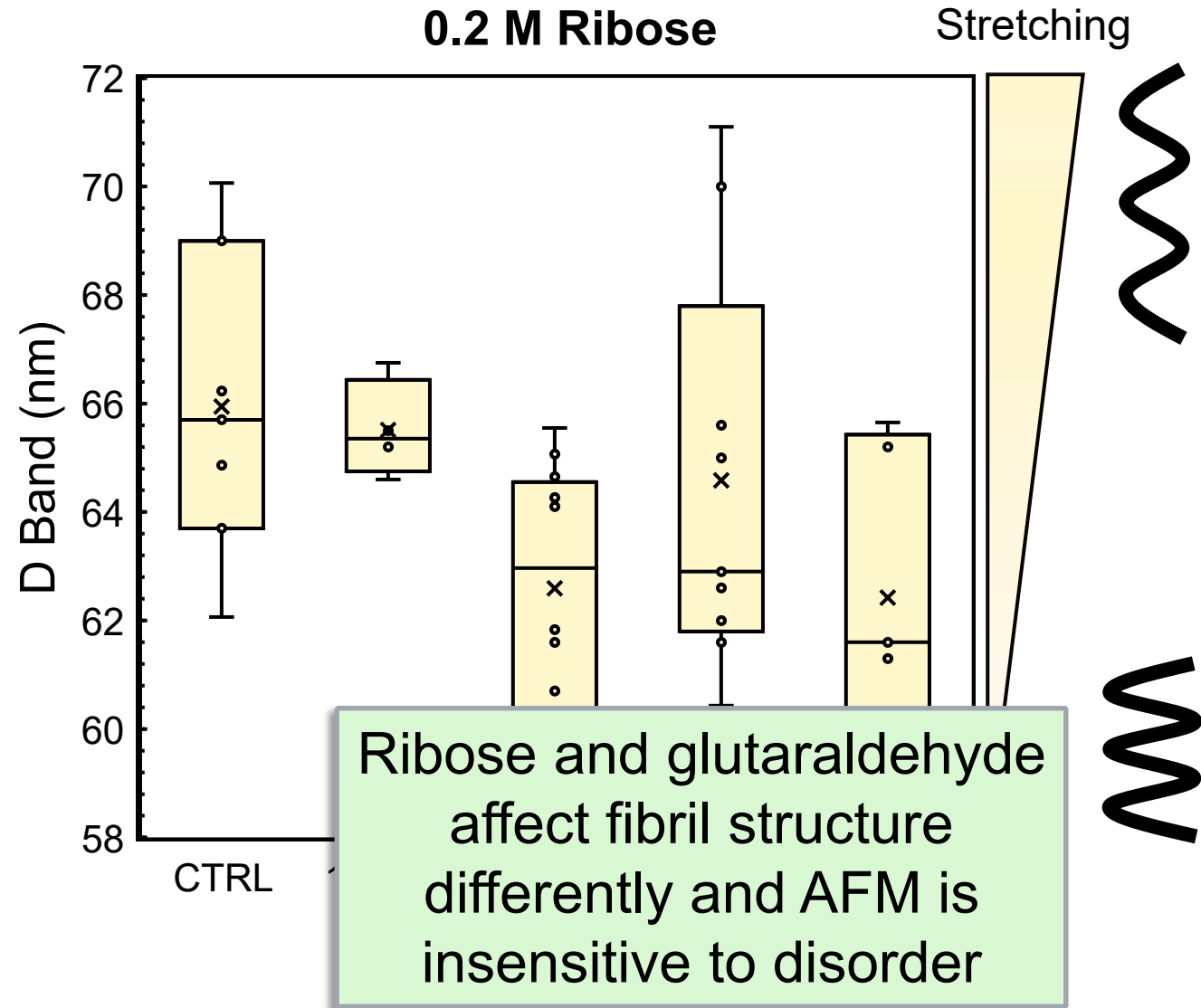
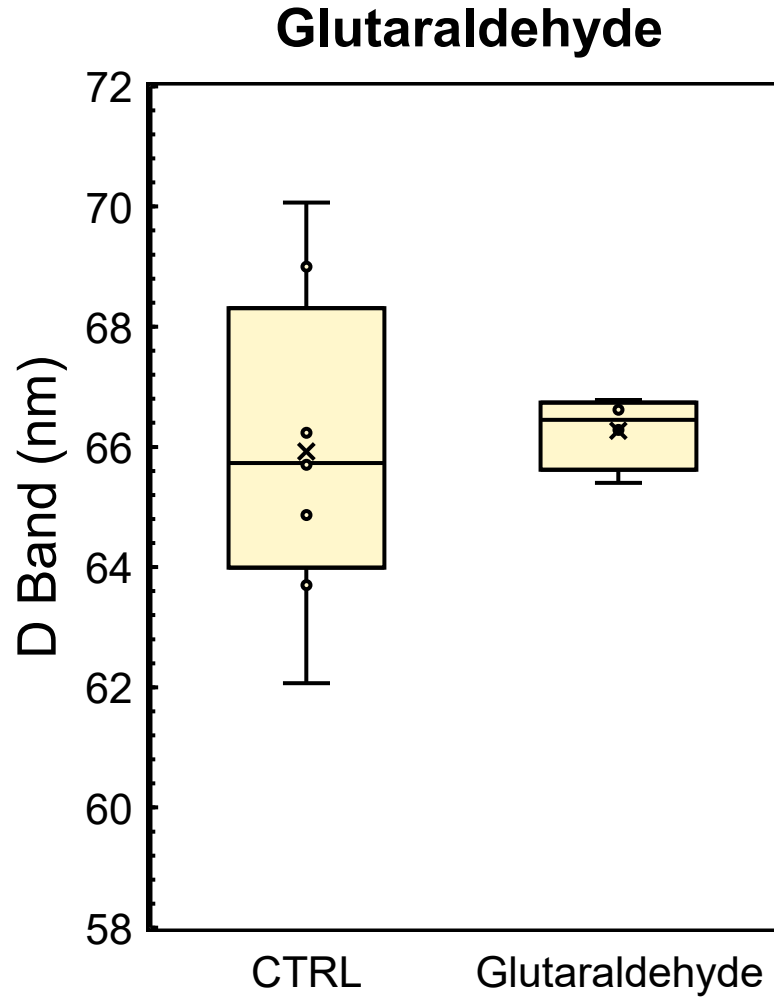
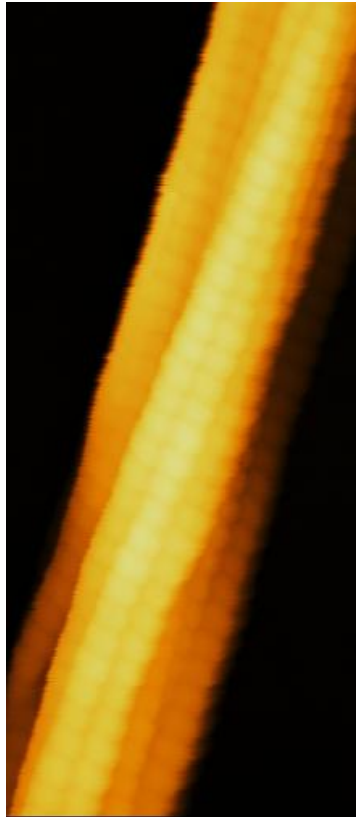
- Fresh *in-vitro* fibrils
- 1, 3, 4, and 6 weeks ribose
- $n = 9$  test and control fibrils
- Extract  $\rho$  value from each fibril, pool test and control, measure differences



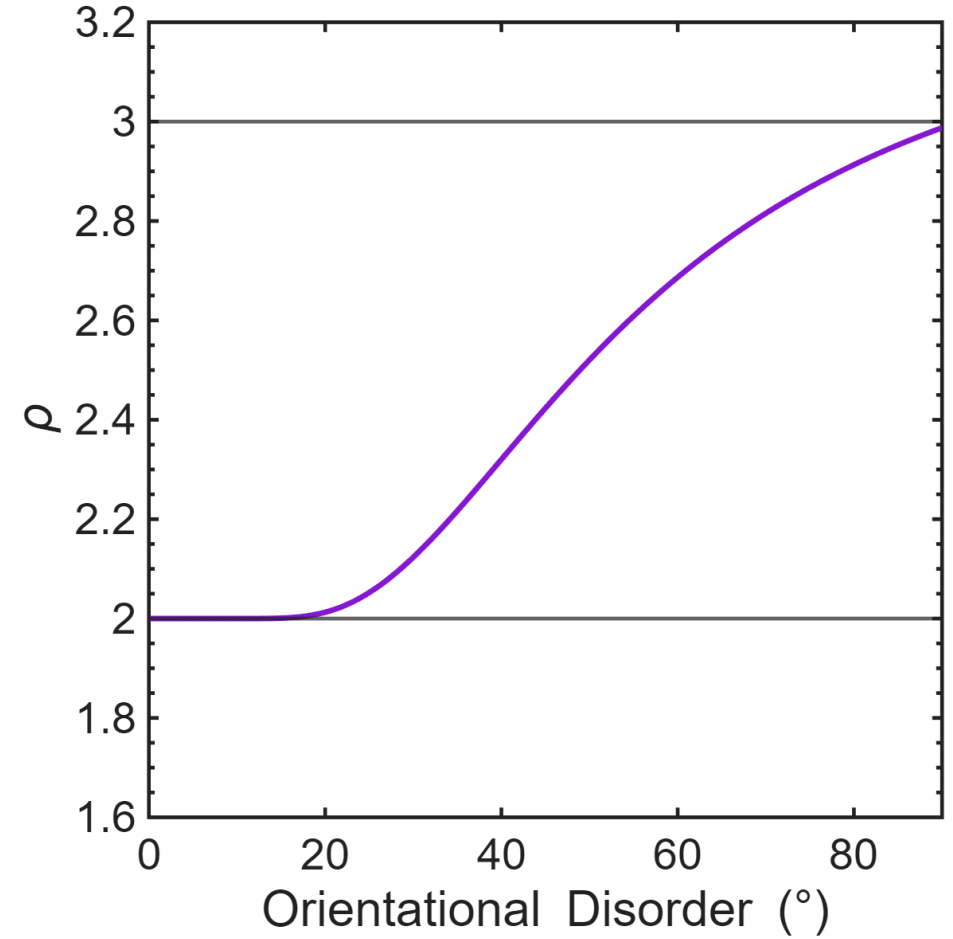
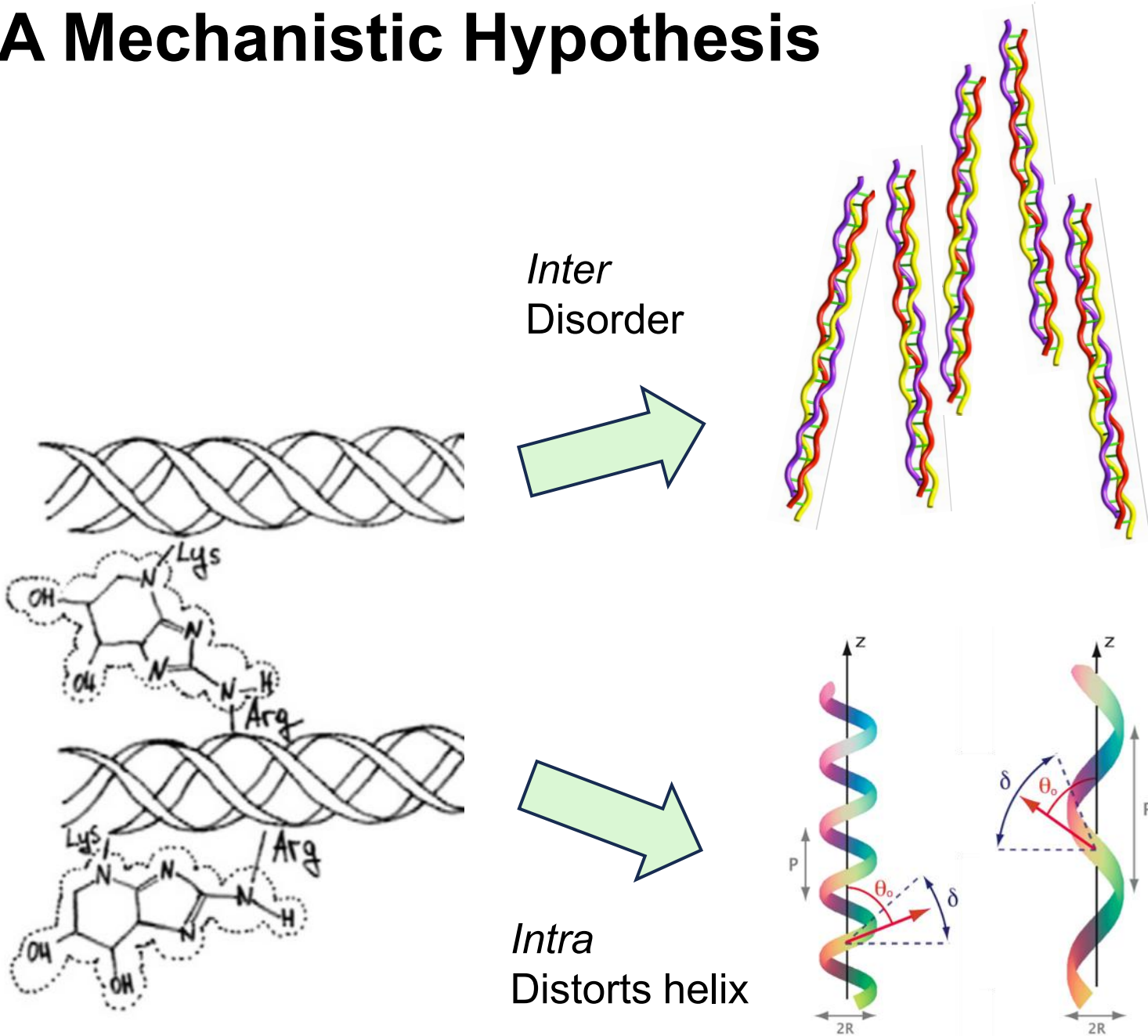
There is a nonlinear change in  $\rho$  with ribose incubation



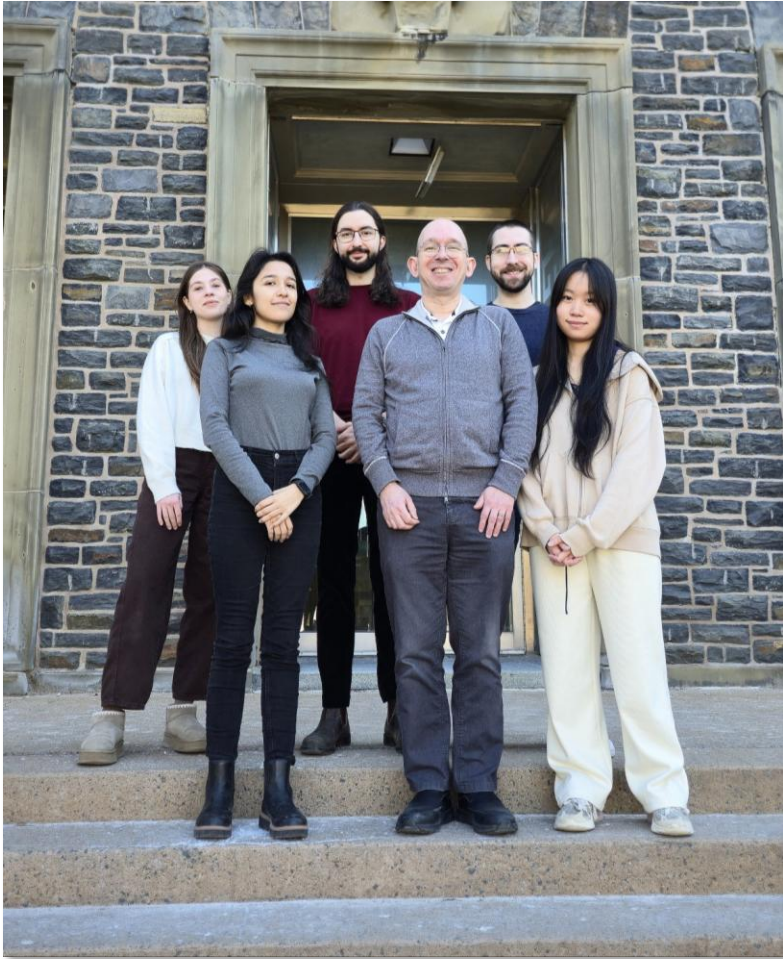
# What about the D Band?



# A Mechanistic Hypothesis



# Acknowledgements



- **Dr. Laurent Kreplak**
- **Dr. Danielle Tokarz**
- Dr. Richard Cisek
- Dr. Andrew Rutenberg
- Dr. Sam Veres
- MacAulay Harvey
- Kateryna Topchylo
- Sara Evans



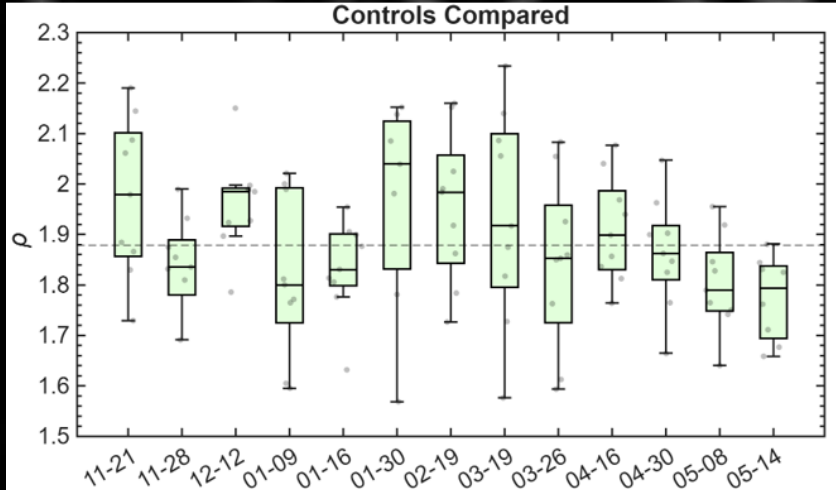
researchNS



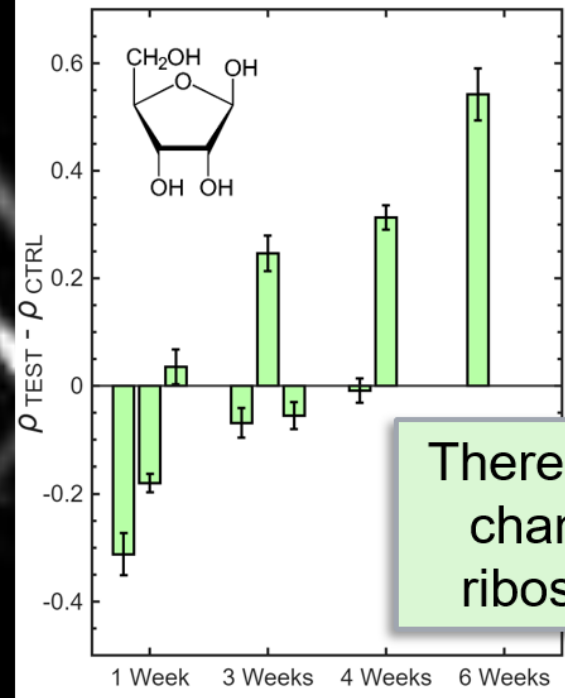
**DALHOUSIE**  
UNIVERSITY



**Saint Mary's**  
University

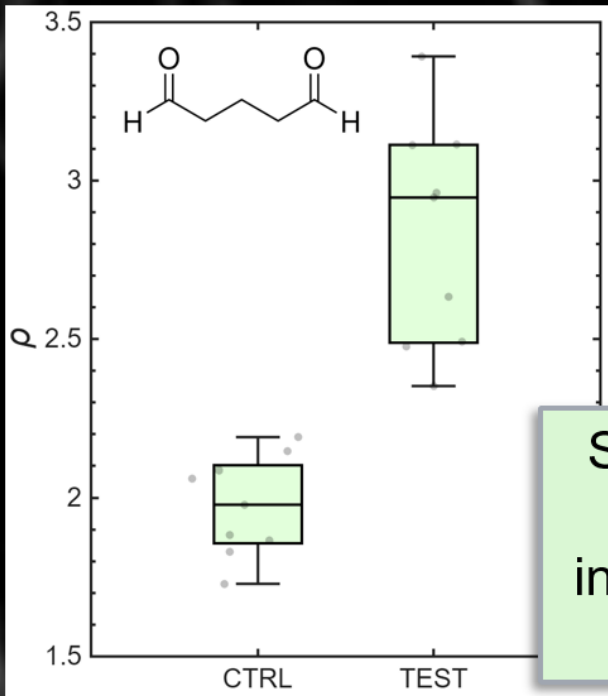


*In-vitro* model for collagen glycation research

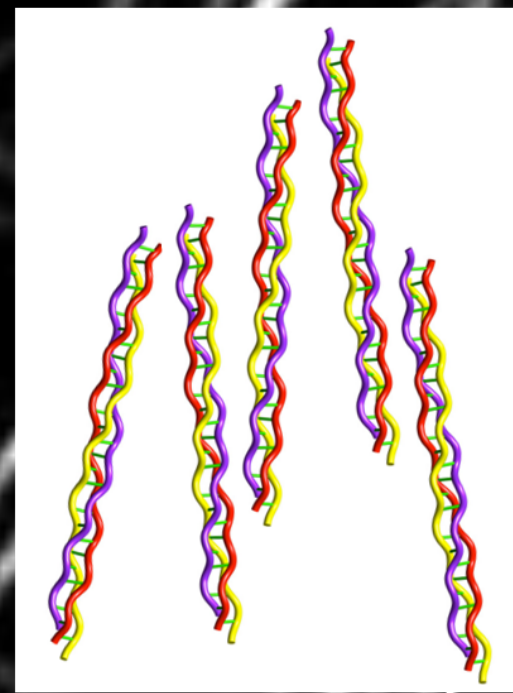


There is a nonlinear change in  $\rho$  with ribose incubation

**Thank You!**

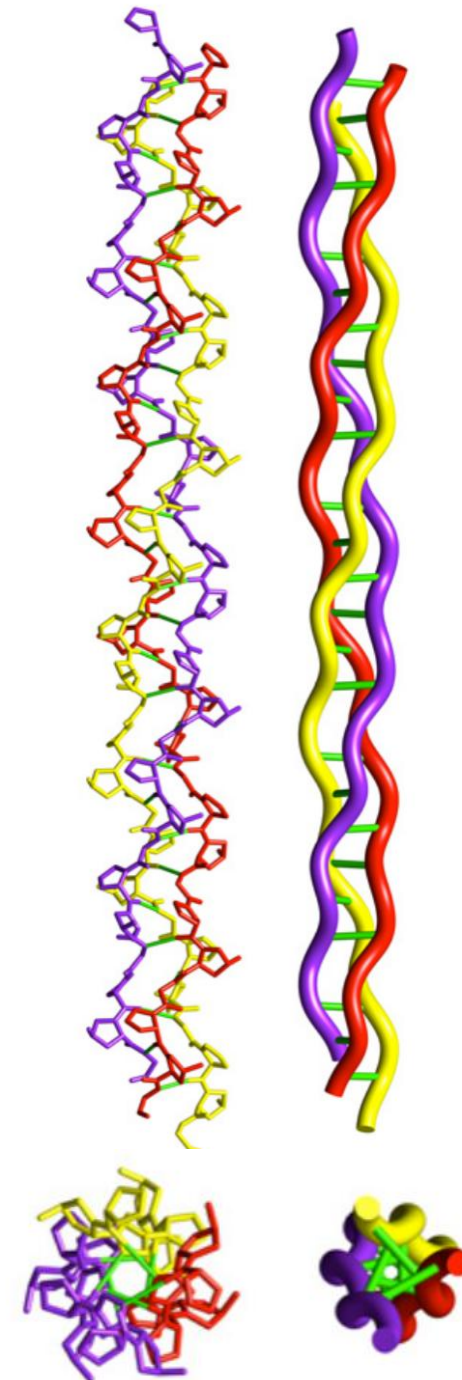
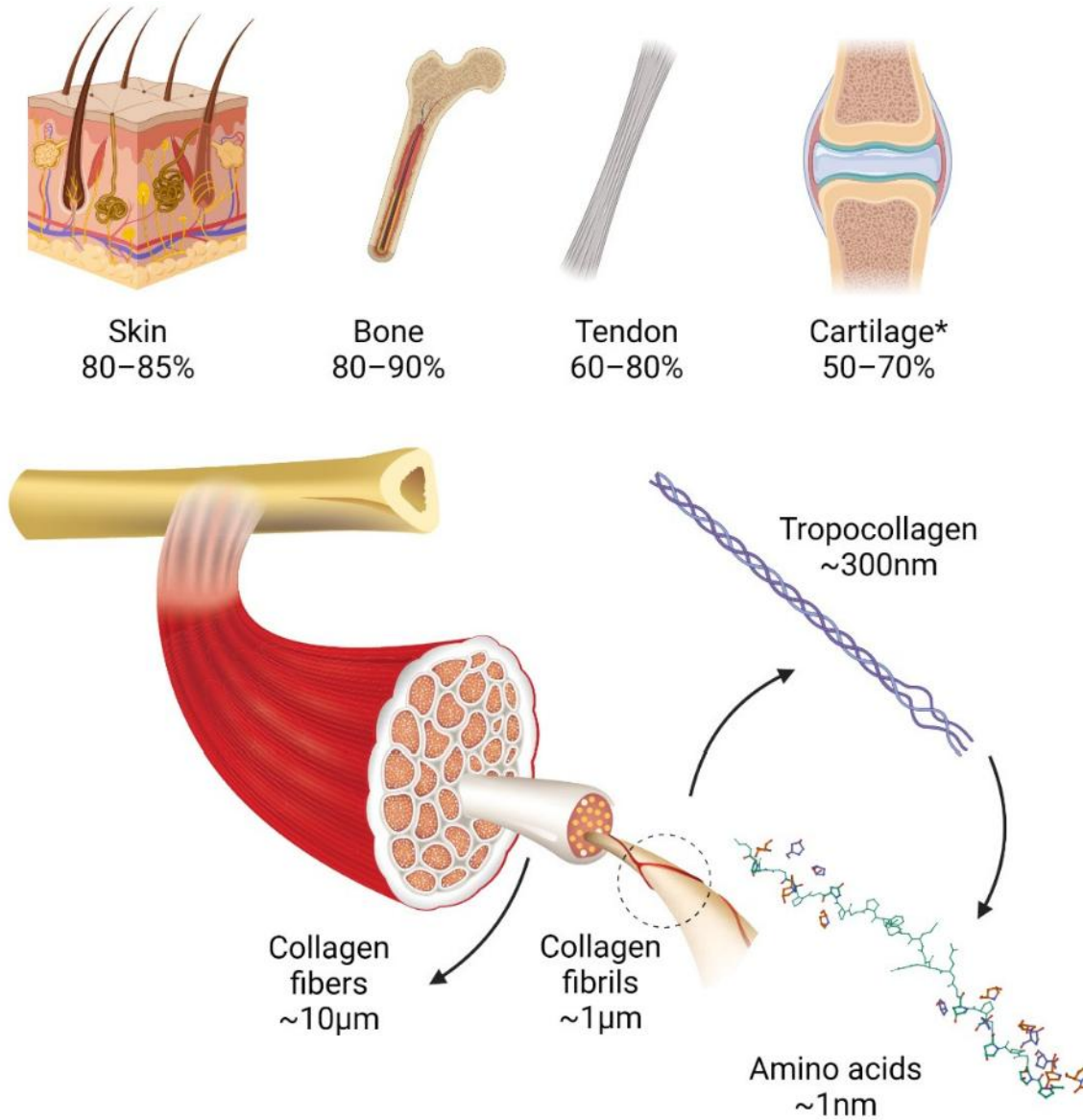


SHG can detect crosslinking-induced changes in collagen



Glycation could cause orientational disorder

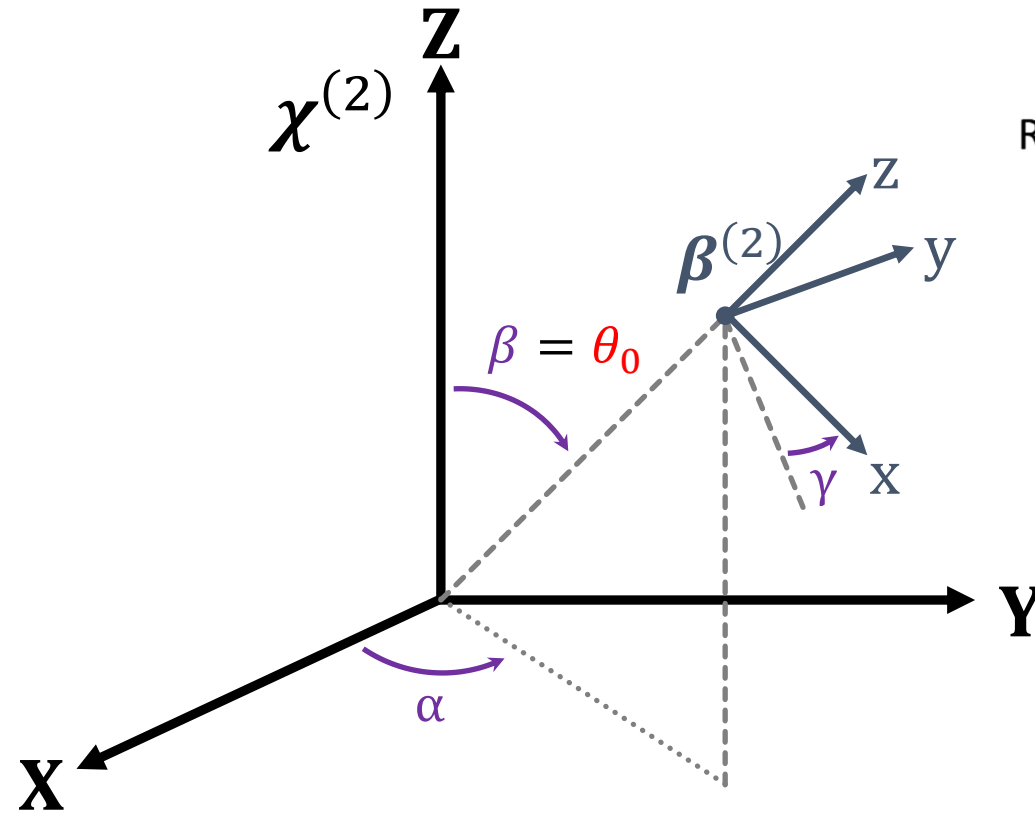
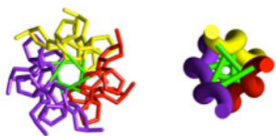
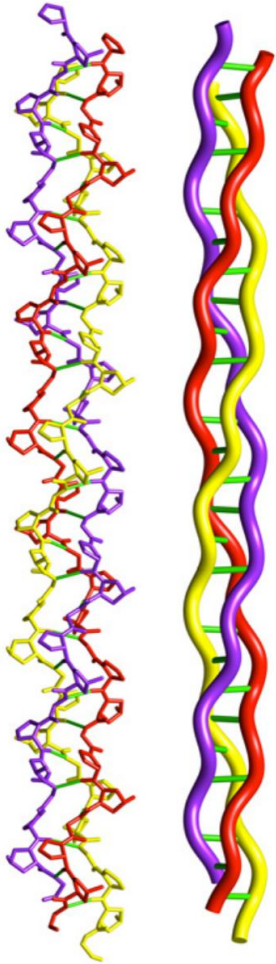
# Collagen



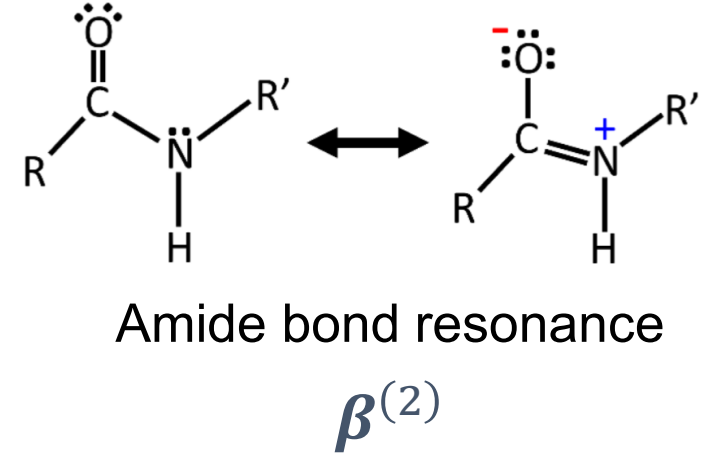
# The Coordinates of the Collagen SHG Signal

Collagen triple helix structure

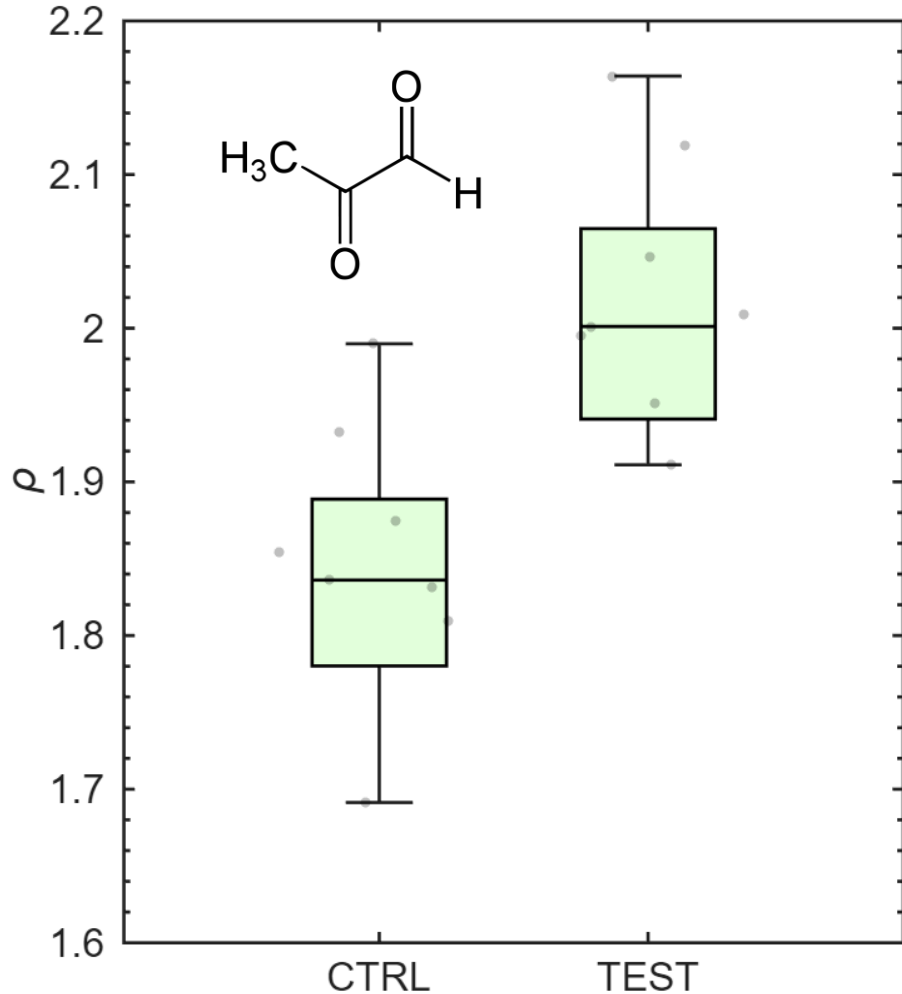
$\chi^{(2)}$



Coordinate system for collagen SHG



# Methylglyoxal Replicates Glutaraldehyde Experiment



PBS Control  
 $n = 9$

Methylglyoxal  
50 mM 24 hours  
 $n = 9$

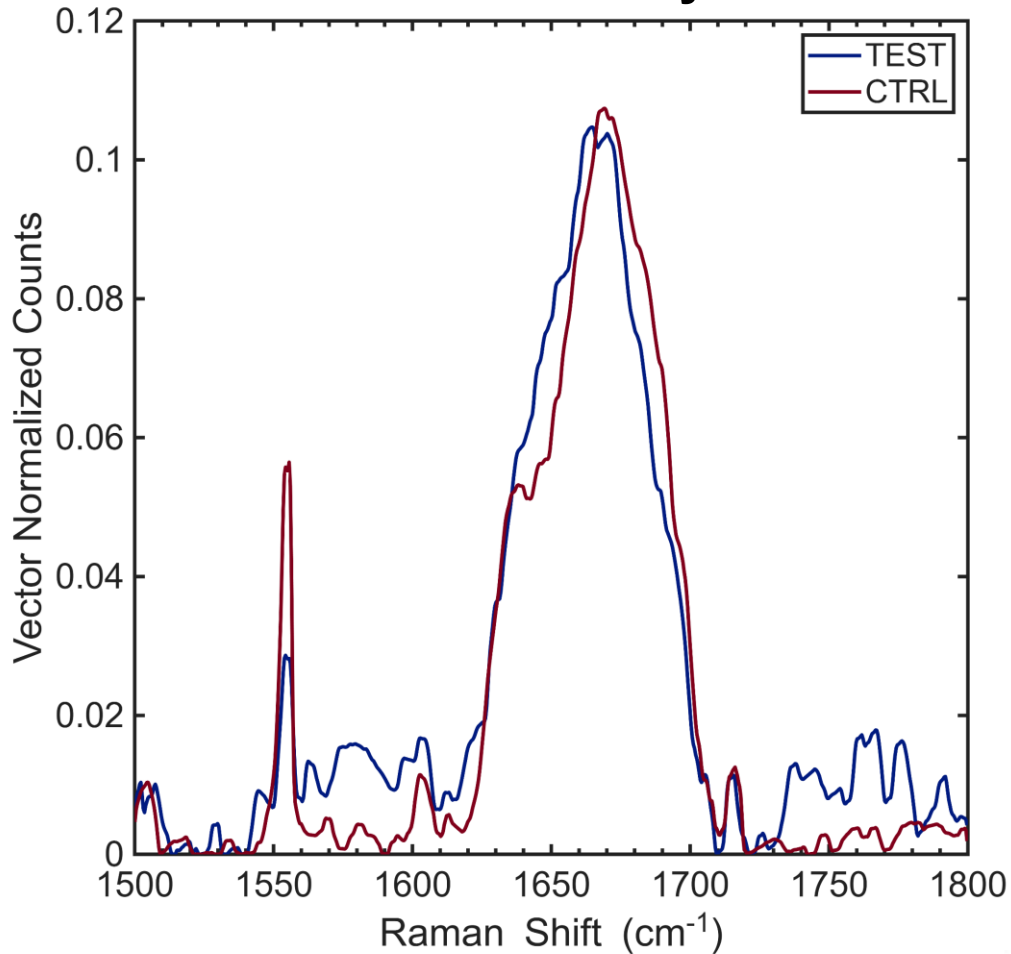
Sample	CTRL $\rho$	TEST $\rho$	$\Delta \rho$	$p$ (T-Test)
GA	2.03	2.88	0.85	0.000019
MGO	1.81	1.96	0.14	0.059

- Both MGO and glutaraldehyde showed an **increase** in  $\rho$
- MGO showed a **smaller** and **less significant** shift than glutaraldehyde

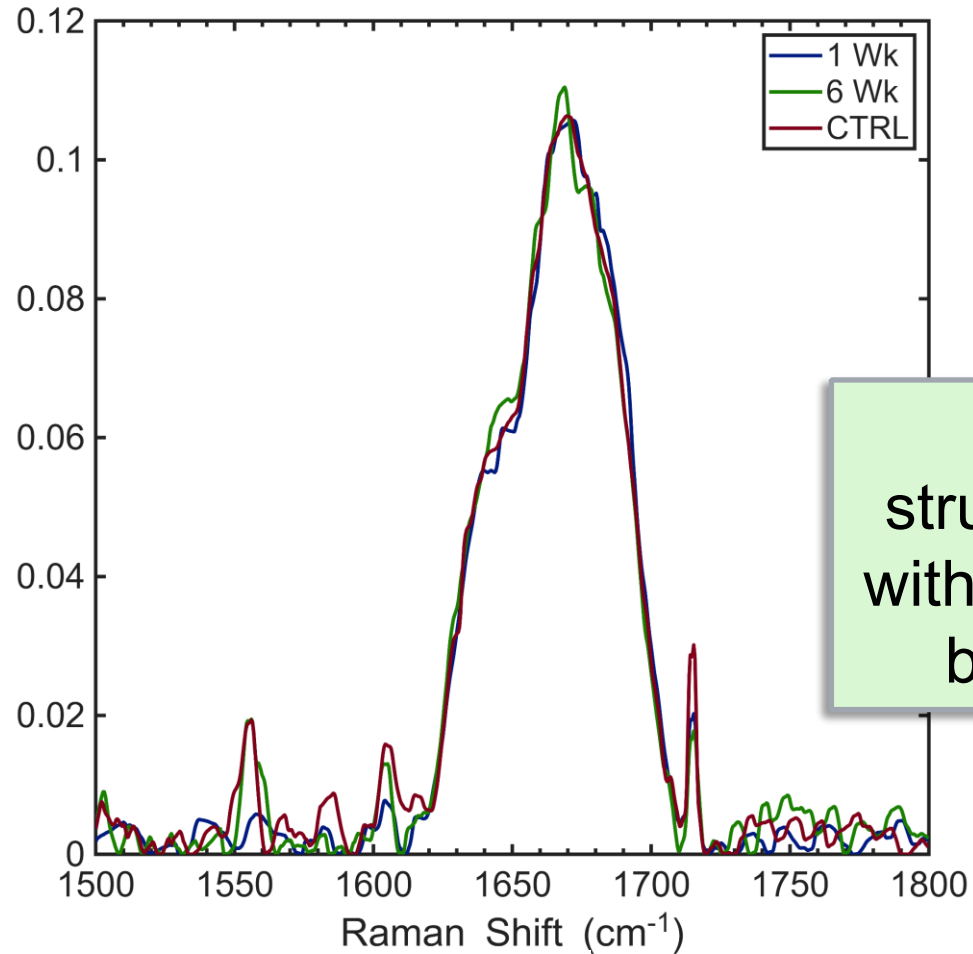
✓ Methylglyoxal repeats the same trend as glutaraldehyde

# Raman Amide I Band Shows Limited Change

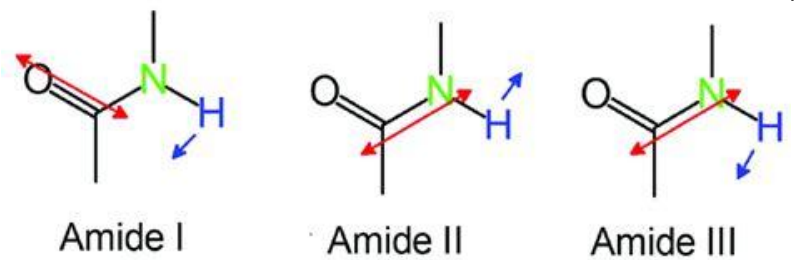
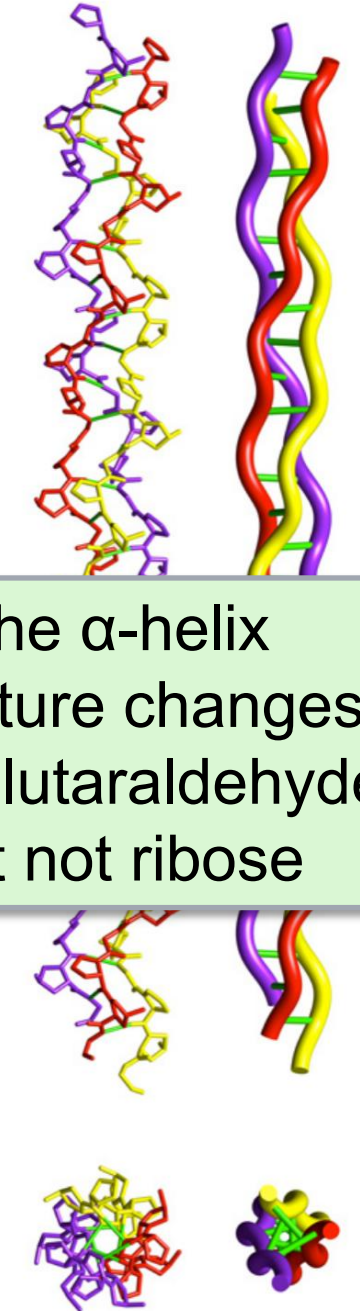
## Glutaraldehyde



## Ribose

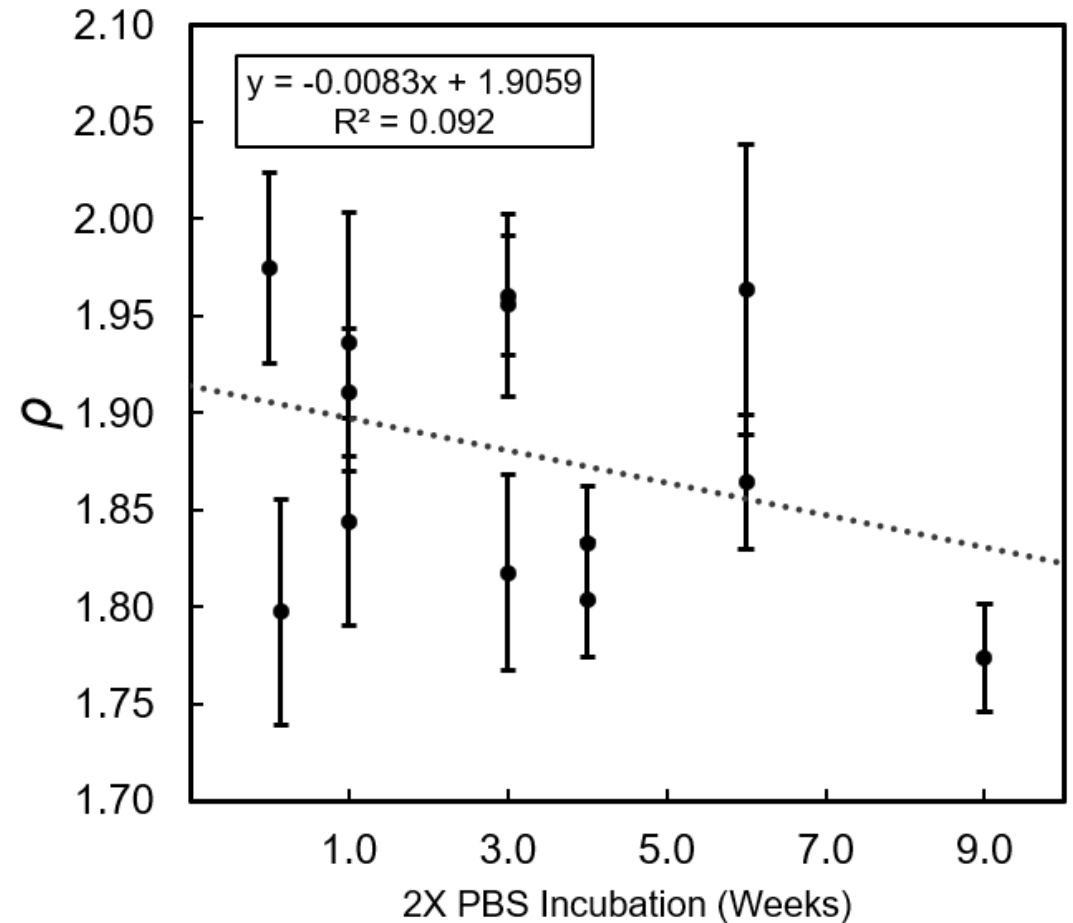
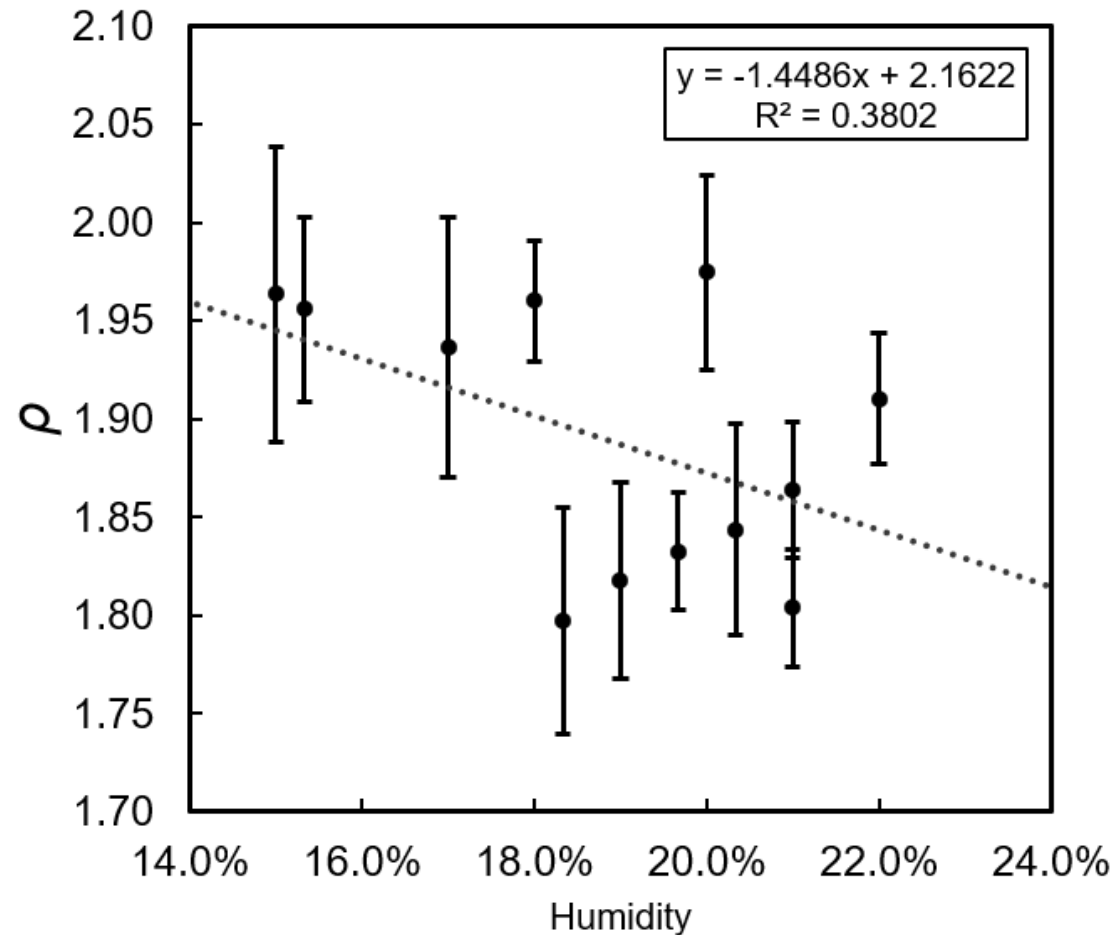


The  $\alpha$ -helix structure changes with glutaraldehyde but not ribose



M. Ichikawa, Micro, (2023)

# Controls are Not Sensitive to Environment

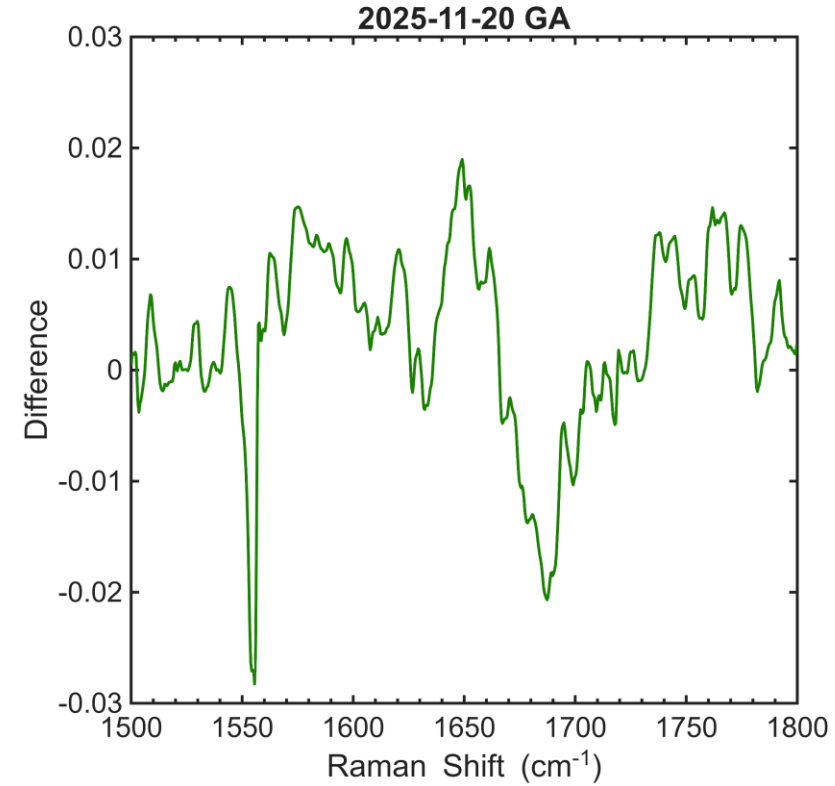
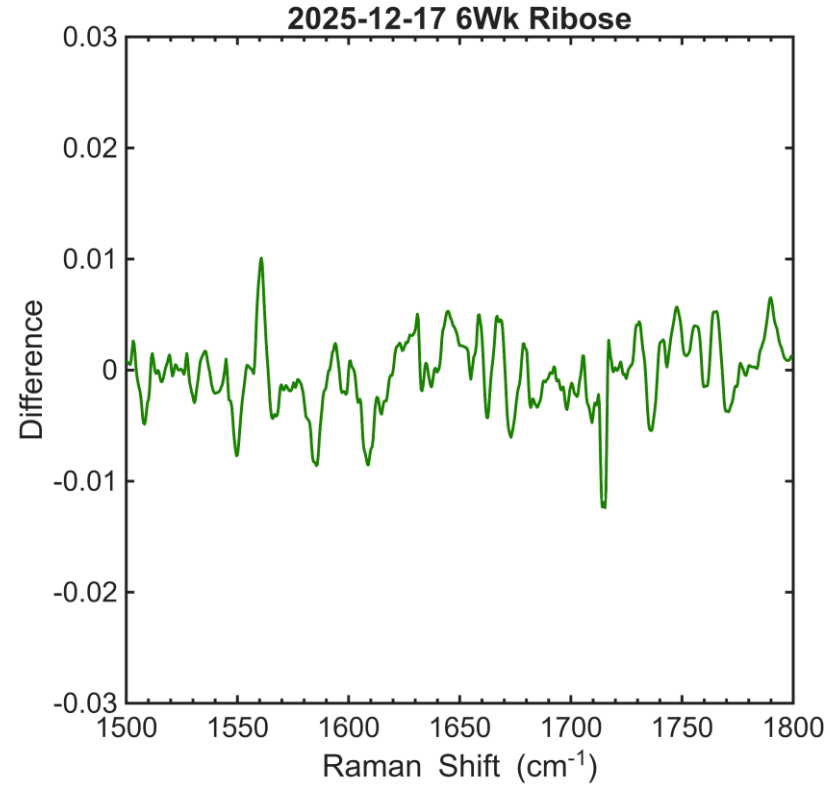
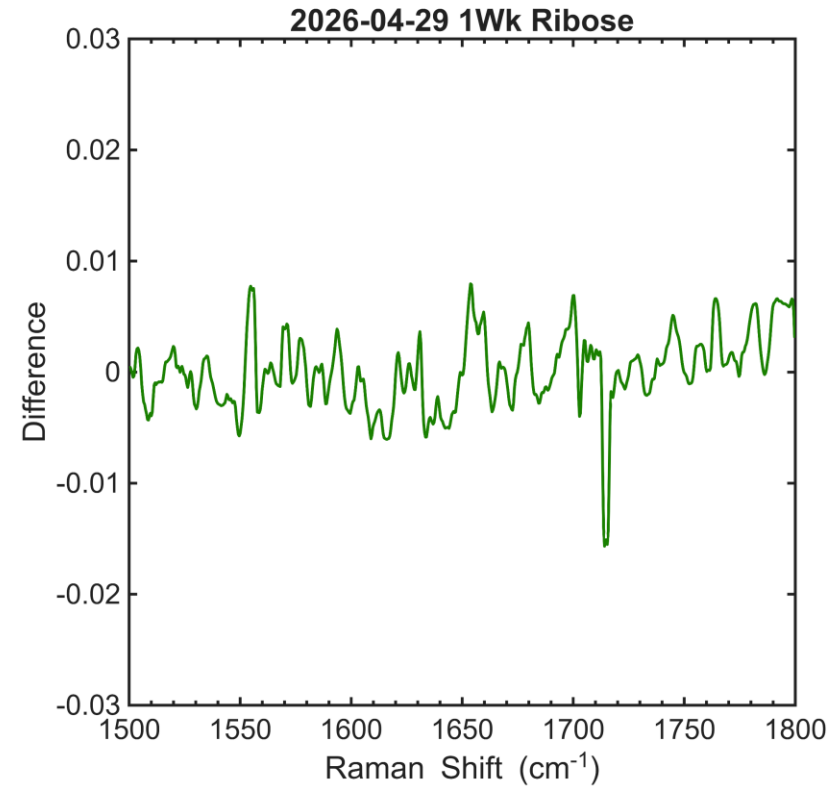


✓ Develop a robust, crosslinking-free in-vitro collagen platform



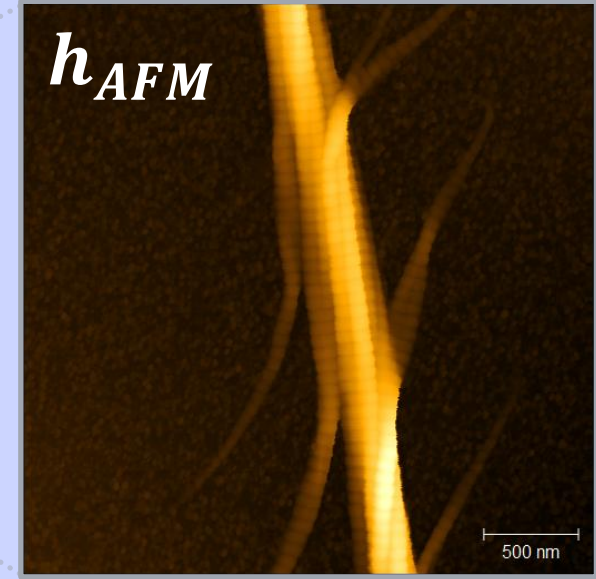
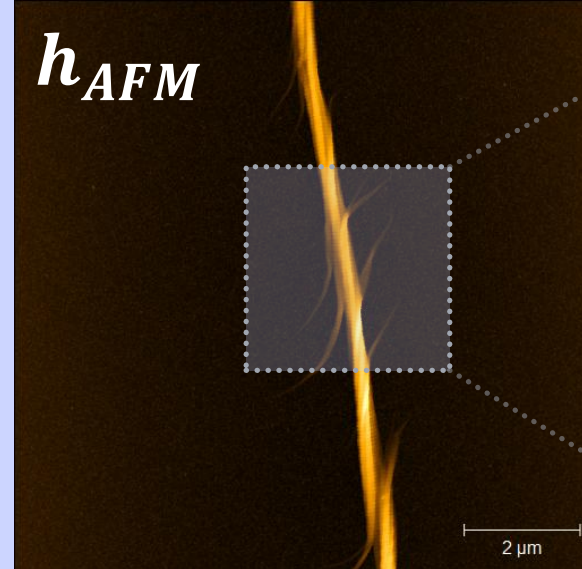
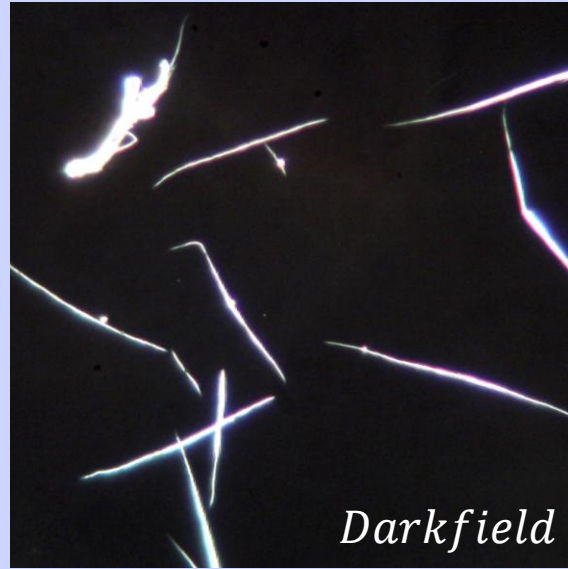
**Thank You!**

# Raman Difference Plots

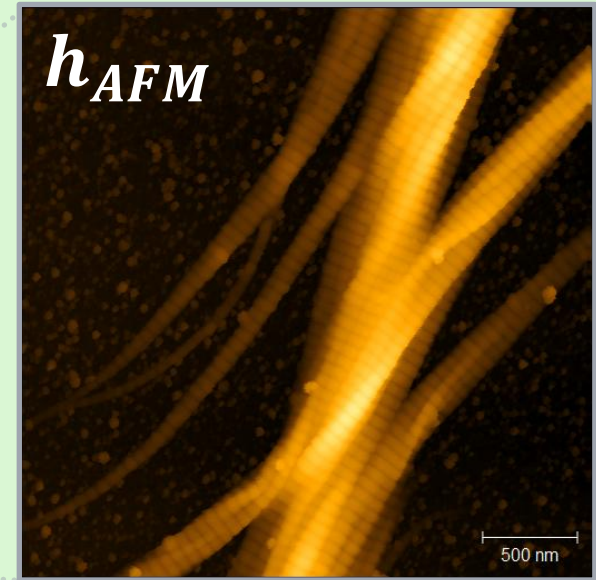
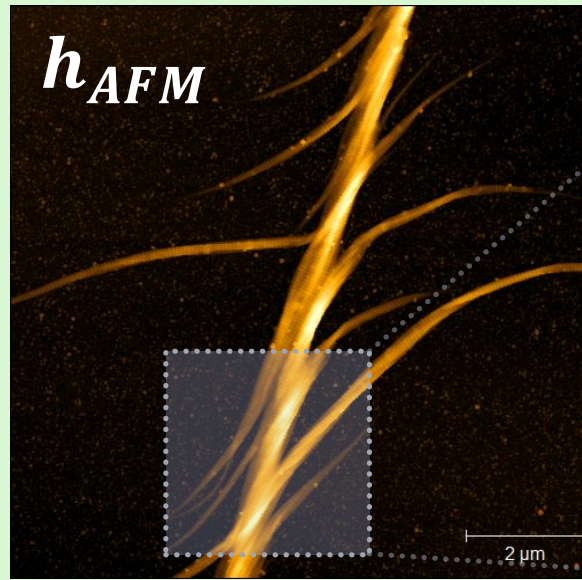


# Fraying of Telocollagen Fibrils Increases with Ribose

**Control**

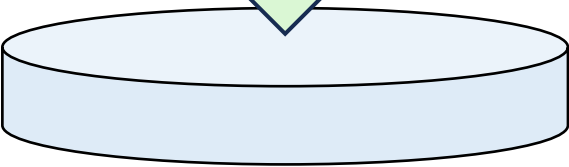
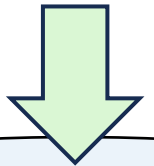


**1 Week  
0.2 M  
Ribose**

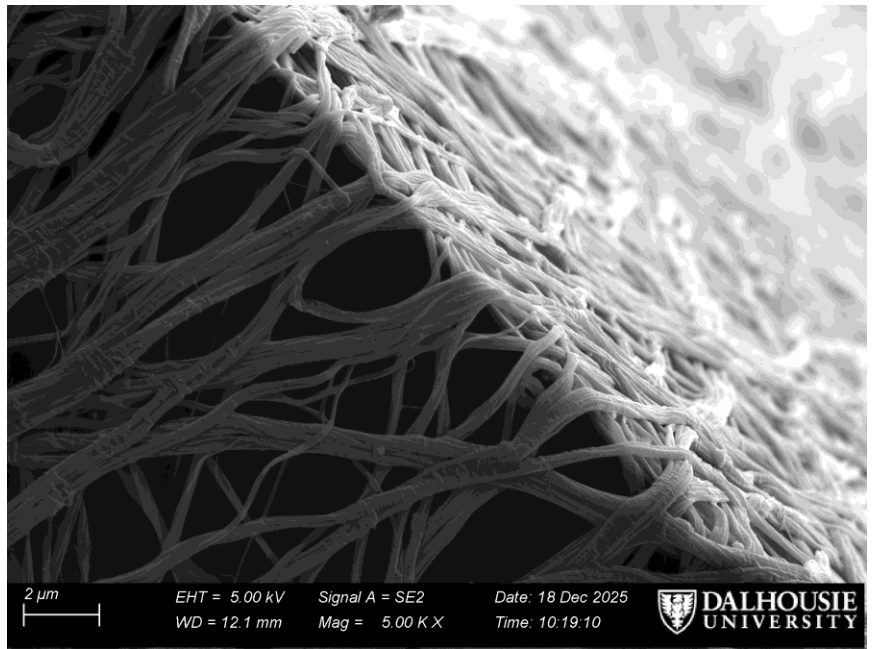
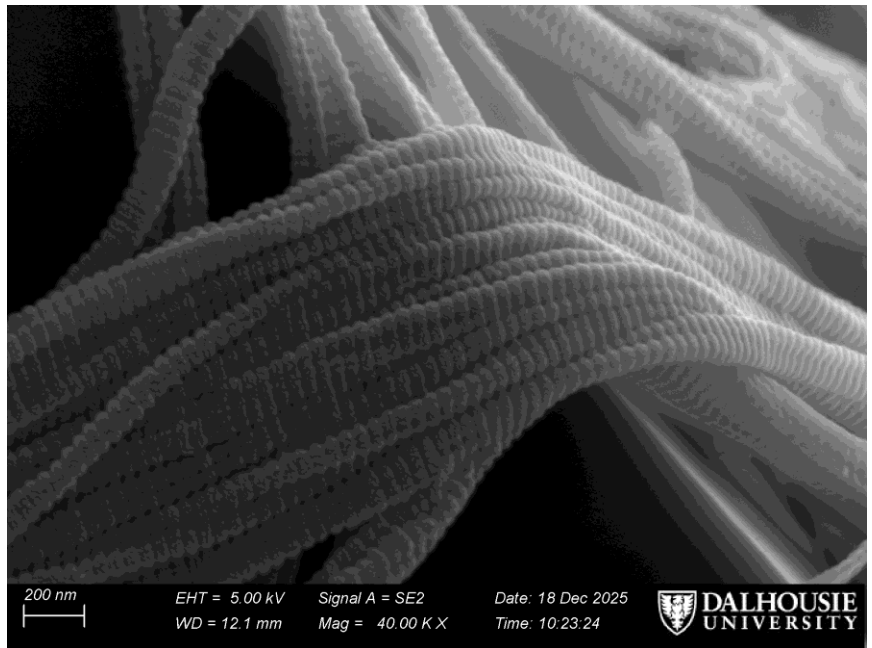
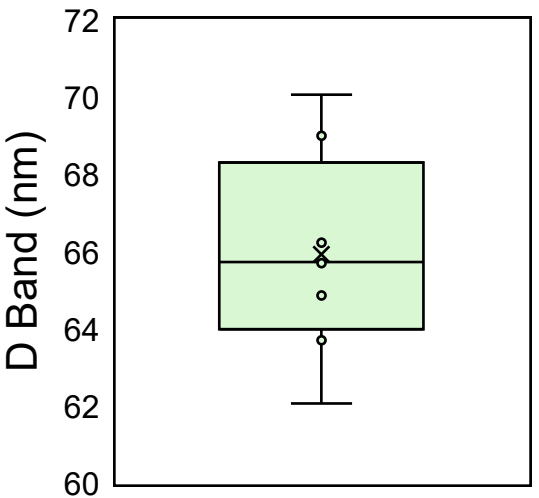
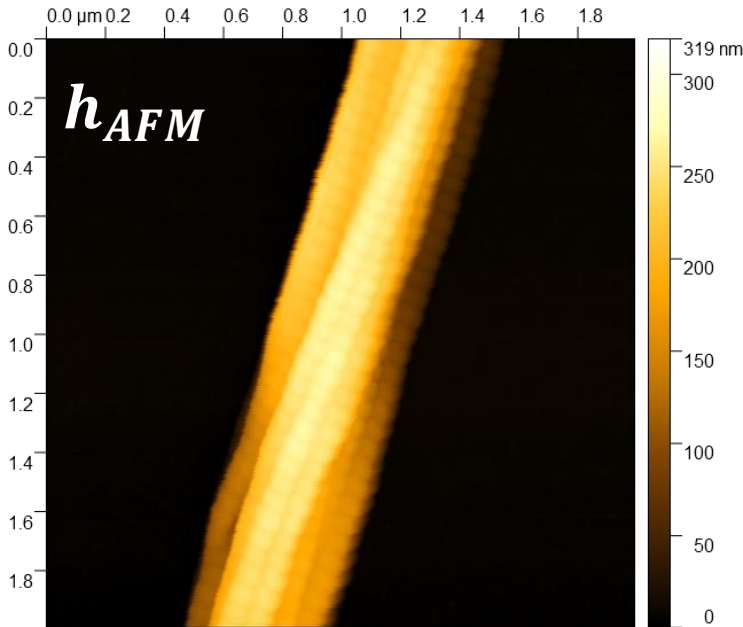
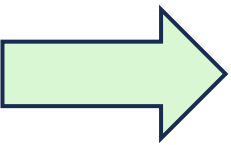


# An *In-Vitro* Fibril Platform

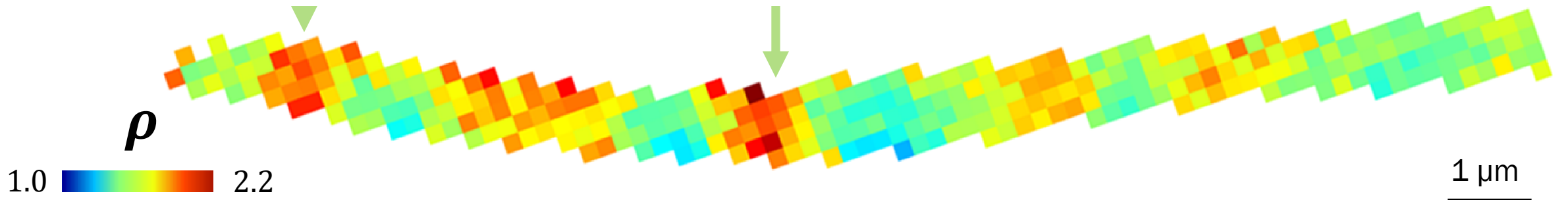
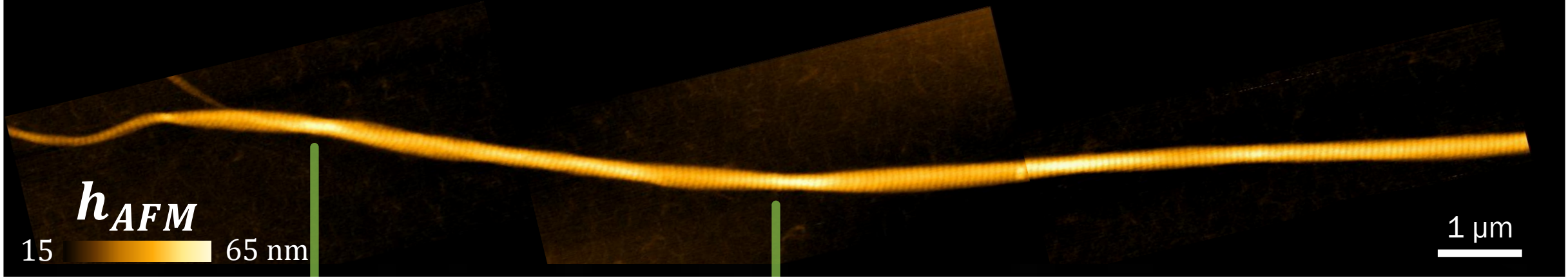
2X PBS  
+  
Bovine  
Telocollagen



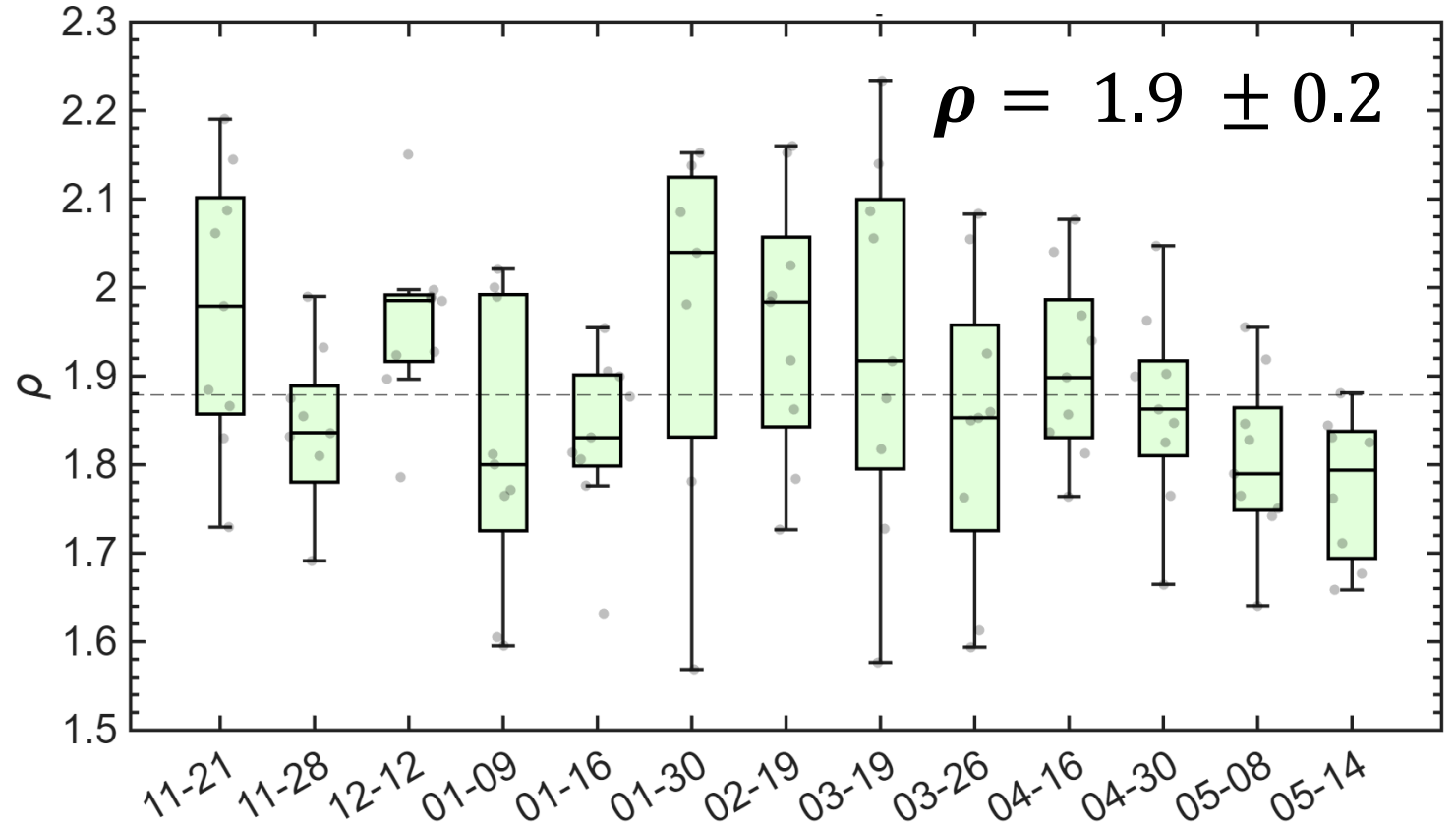
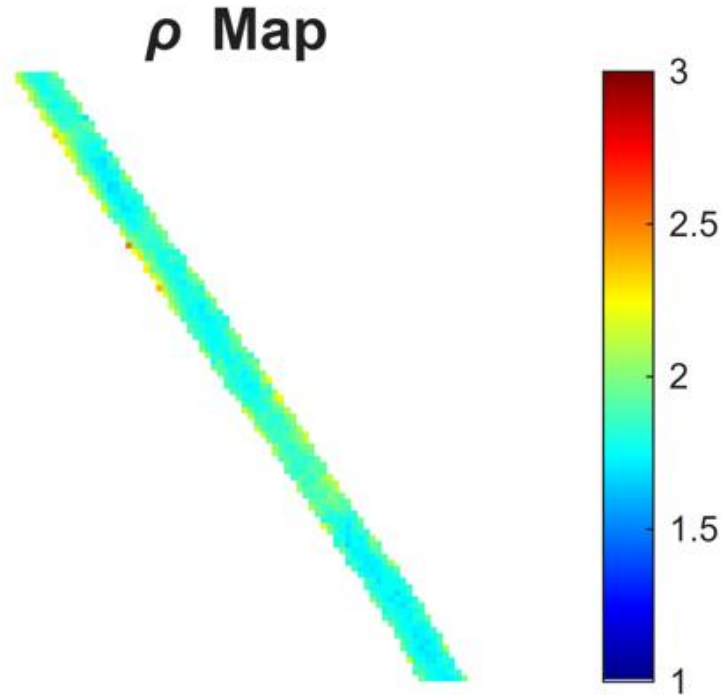
24 hours  
incubation  
at 30°C



# SHG of Collagen Fibrils Reveals Nanoscale Information



# Reproducible Controls Across 13 Experiments



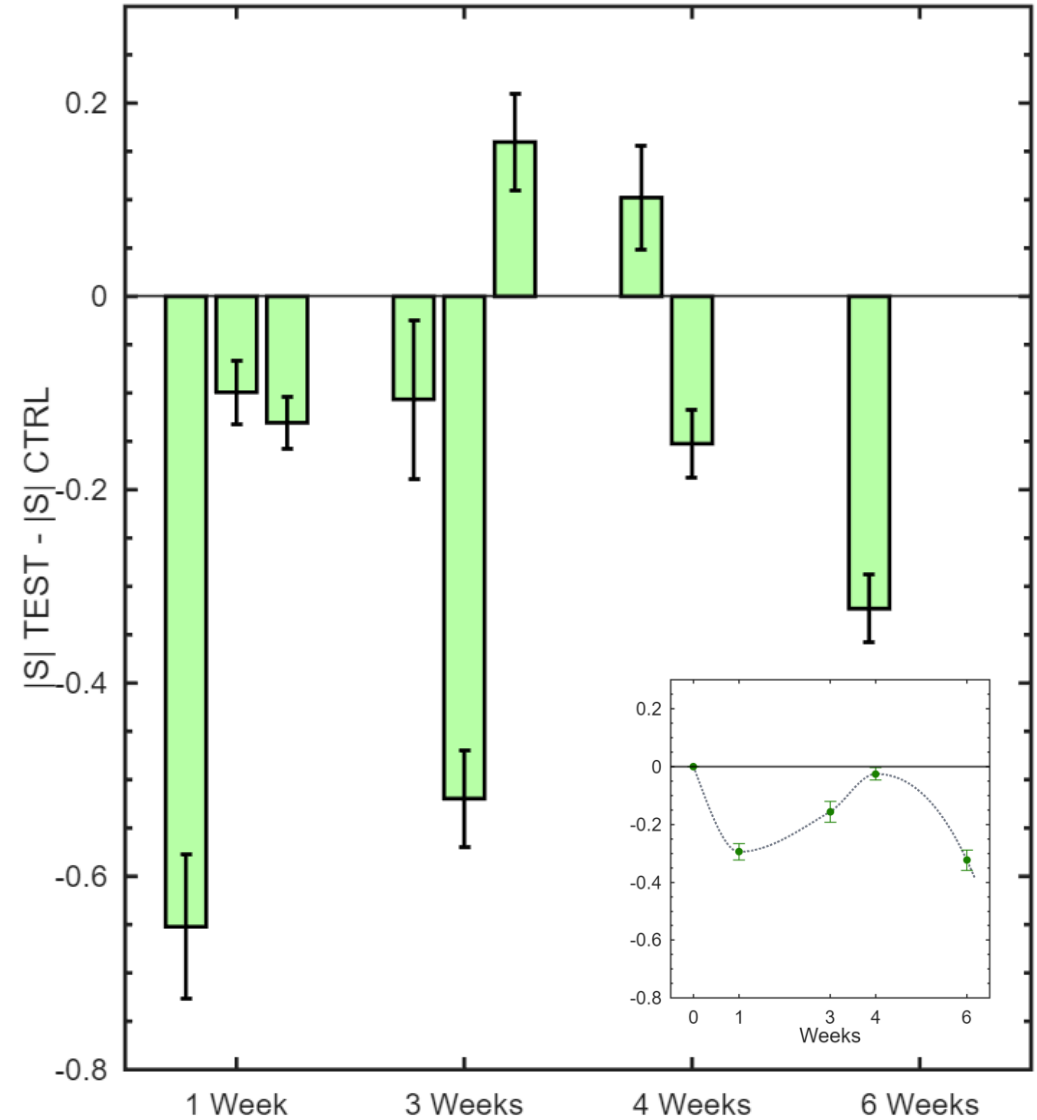
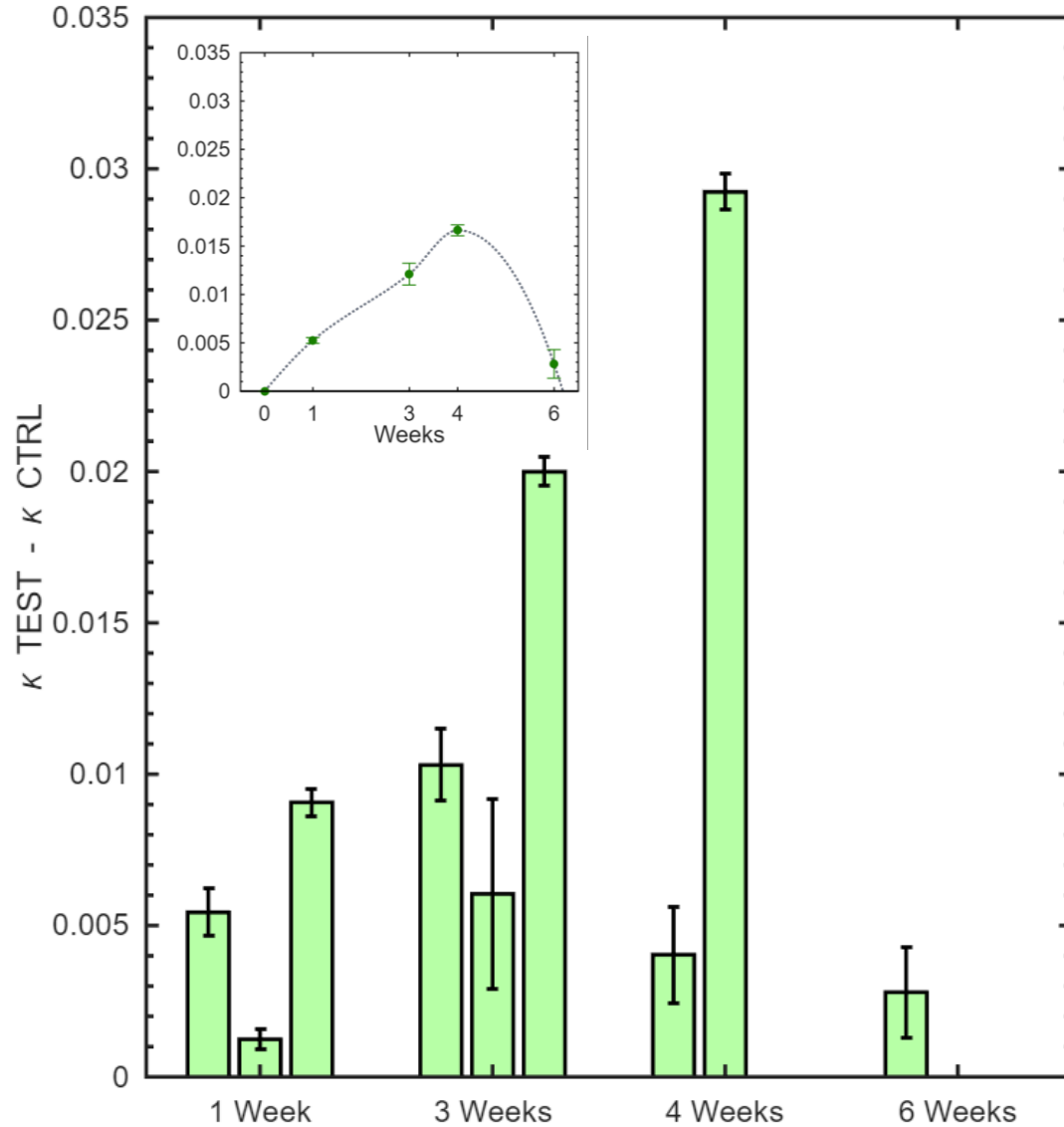
Collagen fibrils tend to have homogenous  $\rho$ , however, there are fibril-to-fibril differences

Due to synthesis variations, a new control is acquired for every experiment

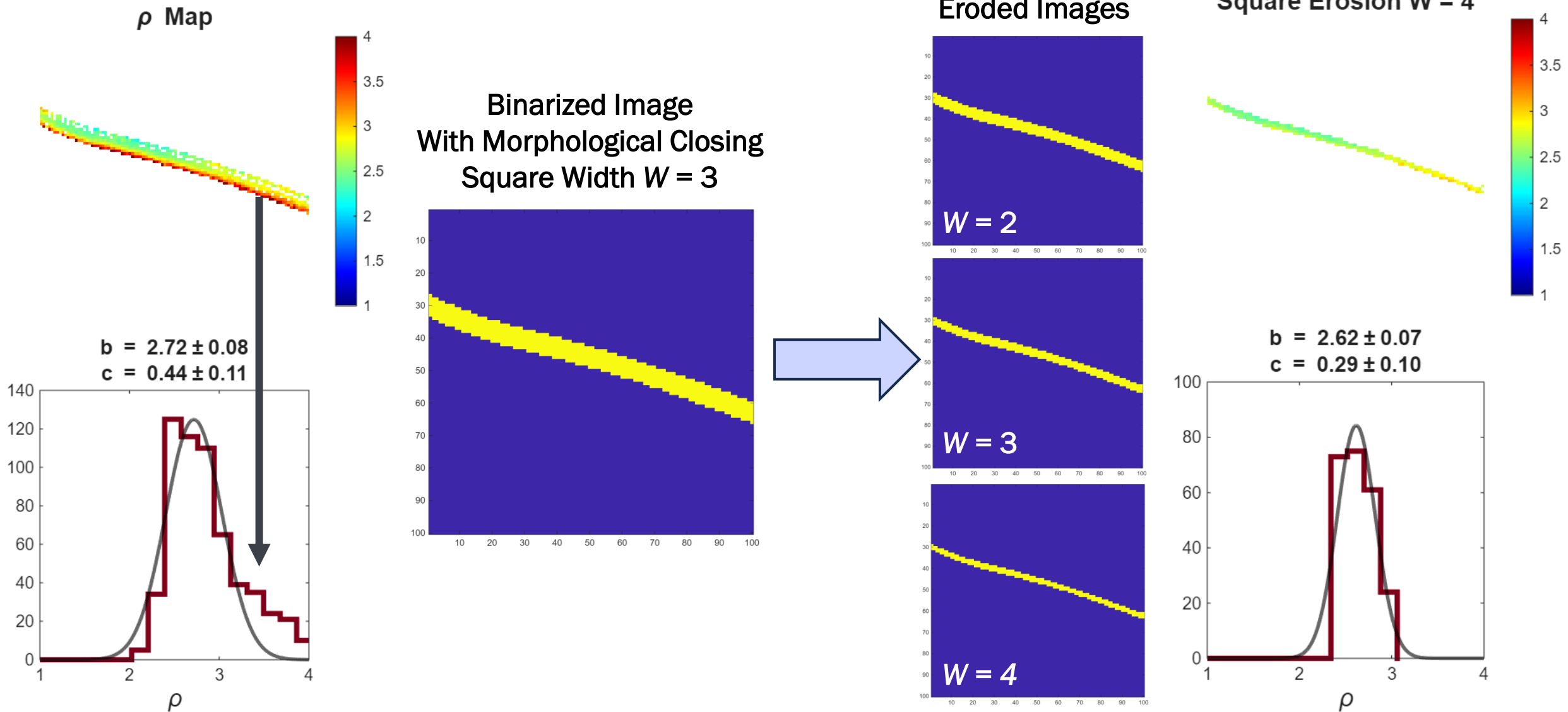
$n = 114$  fibrils measured

# Chiral ( $\kappa$ ) and Asymmetry ( $S$ ) Parameters

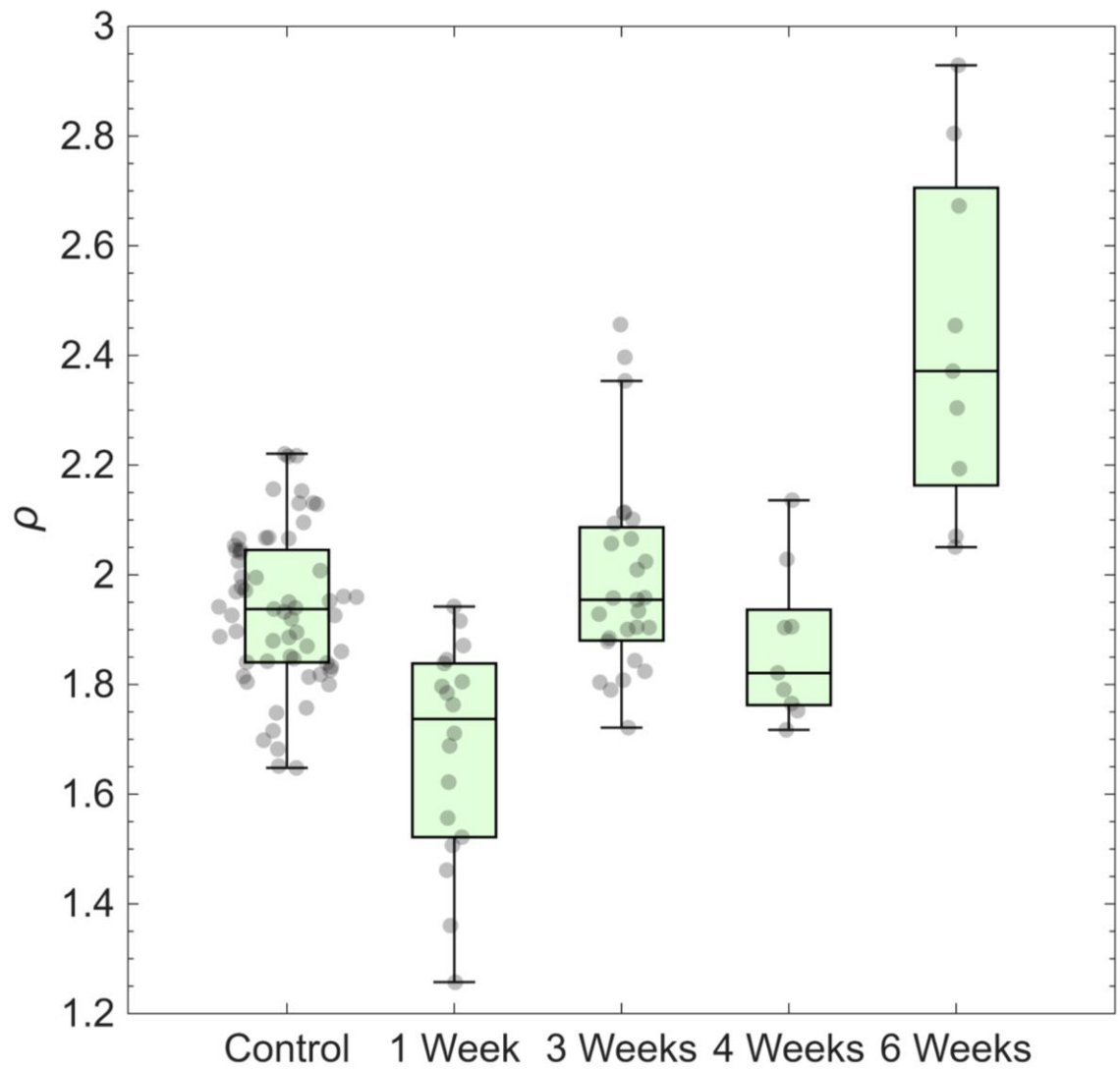
$$\kappa = \frac{\chi_{XYZ}^{(2)}}{\chi_{ZXX}^{(2)}} \quad S = \frac{\chi_{XXX}^{(2)}}{\chi_{ZXX}^{(2)}}$$



# Removing Outlier Pixels with Erosion Filtering



# Alternative Ribose Plots



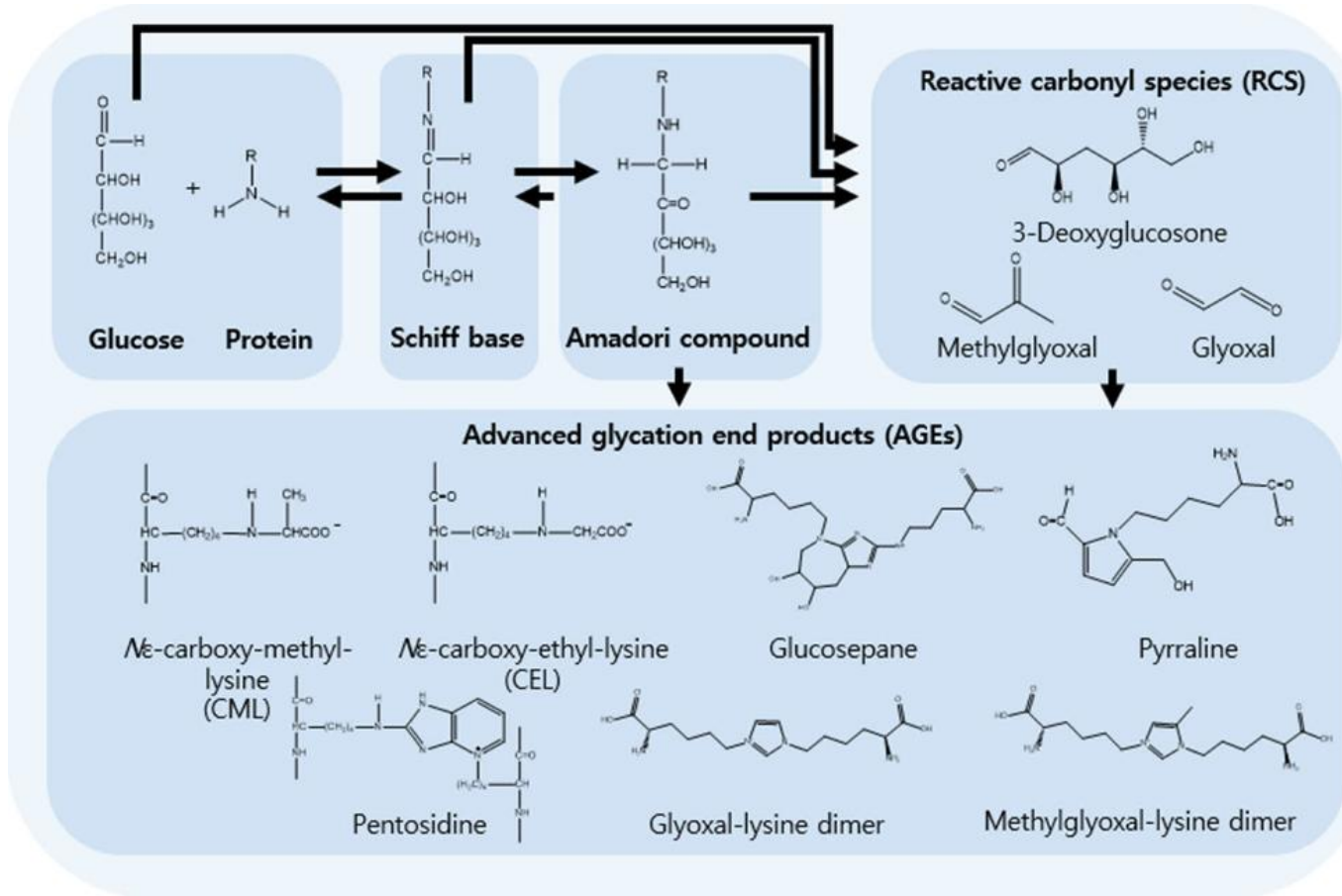
# Conclusions

- We developed a robust *in-vitro* model for collagen SHG
- SHG is highly sensitive to crosslinking in collagen fibrils
- Molecular disorder increases in glycated collagen fibrils
- Ribose and glutaraldehyde affect collagen structure differently
- The glycation process is rich, multifaced, and nonlinear

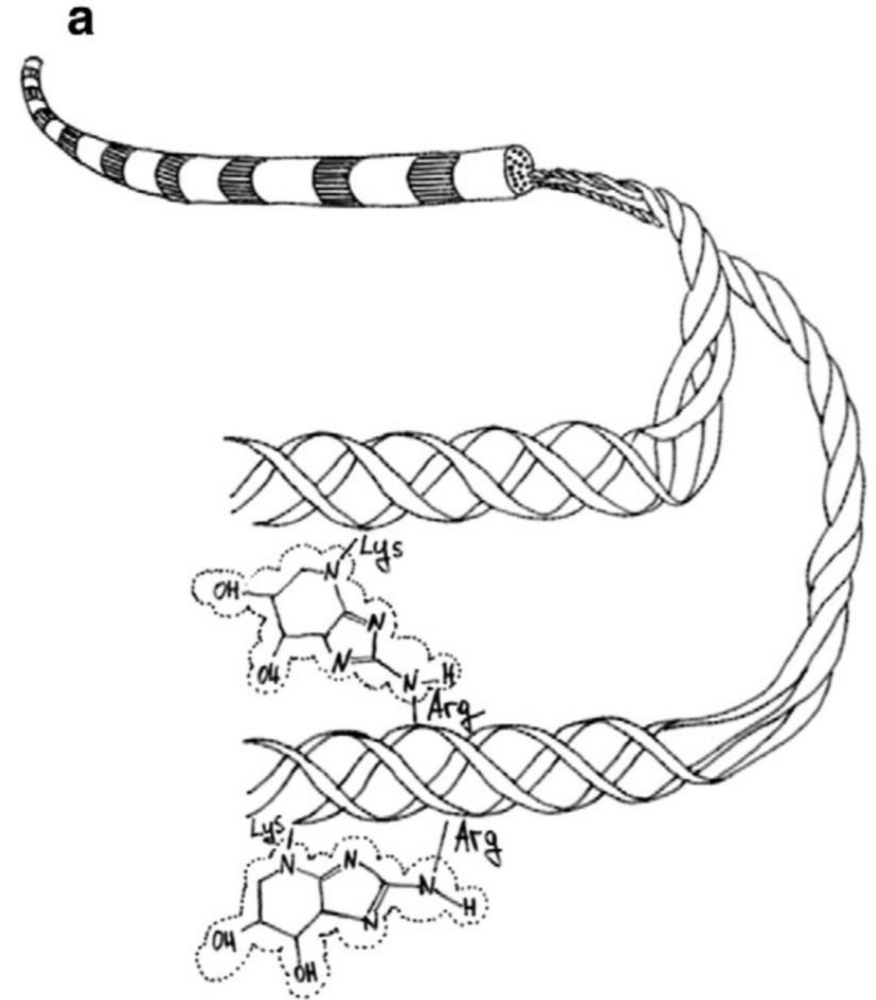
# Future Directions

- Additional ribose experiments
- Further glutaraldehyde experiments to distinguish between *intra-* and *inter-*molecular crosslinking
- Additional measurements with fluorescence anisotropy and X-ray diffraction to test the disorder hypothesis
- Exploring further SHG and polarization parameters

# Glycation Chemistry of Collagen Fibrils



S. Cho et. al., Reviews in Analytical Chemistry, (2022).



A. Gautieri et. al., Matrix Biology, (2014)

# Summary – What Did We Observe?

## Glutaraldehyde

1 experiment with 50 mM, 1 hr

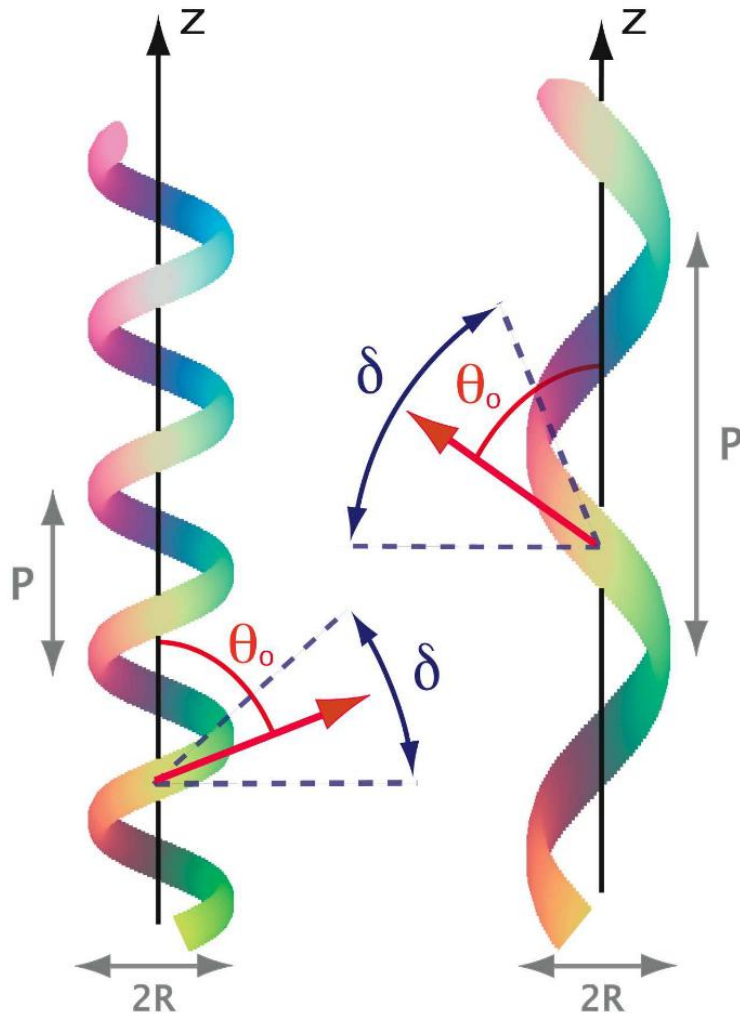
- Pitch angle **decreases** to 40°
- Width of angles **increases**
- D-band remains the **same**

## Ribose Time Series

7 experiments with 0.2 M and 1, 3, 4, and 6 weeks

- Pitch angle **decreases** to 42 – 44°
- Width of angles may **increase** at 6 weeks ribose
- Nonlinear trend, **increase** in pitch at 1 week
- D-band **decreases** with incubation time

# Using PIPO-SHG to Find Helical Pitch Angle



Tiaho et. al., Optics Express, (2007)

$$\rho = \frac{\chi_{ZZZ}^{(2)}}{\chi_{ZXX}^{(2)}} \quad \cos^2 \theta_0 = \frac{\rho}{\rho + 2}$$

$$\rho = 3.0 \rightarrow \langle \theta_0 \rangle = 39.2^\circ$$

$$\rho = 2.0 \rightarrow \langle \theta_0 \rangle = 45.0^\circ$$

$$\sigma_\rho \sim 0.05 \rightarrow \sigma_{\langle \theta_0 \rangle} < 1^\circ$$

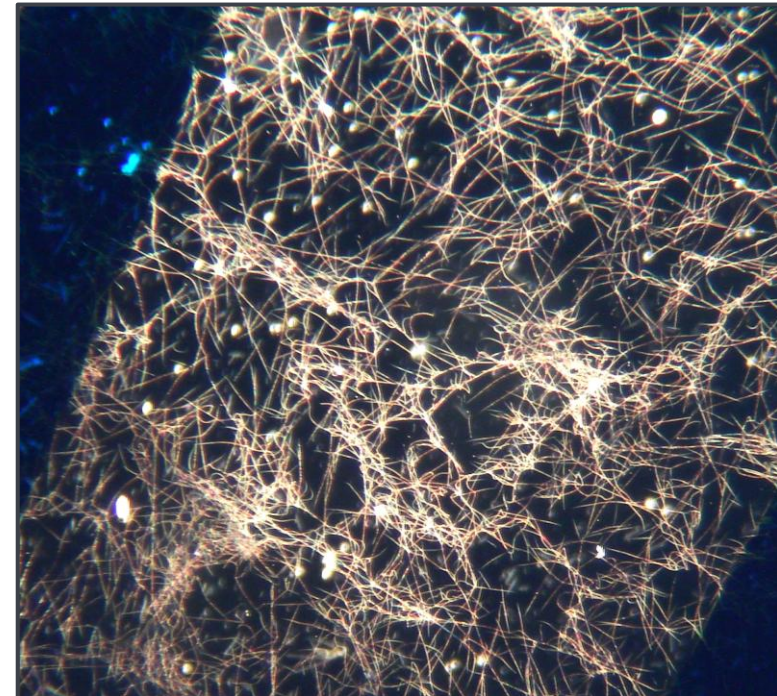
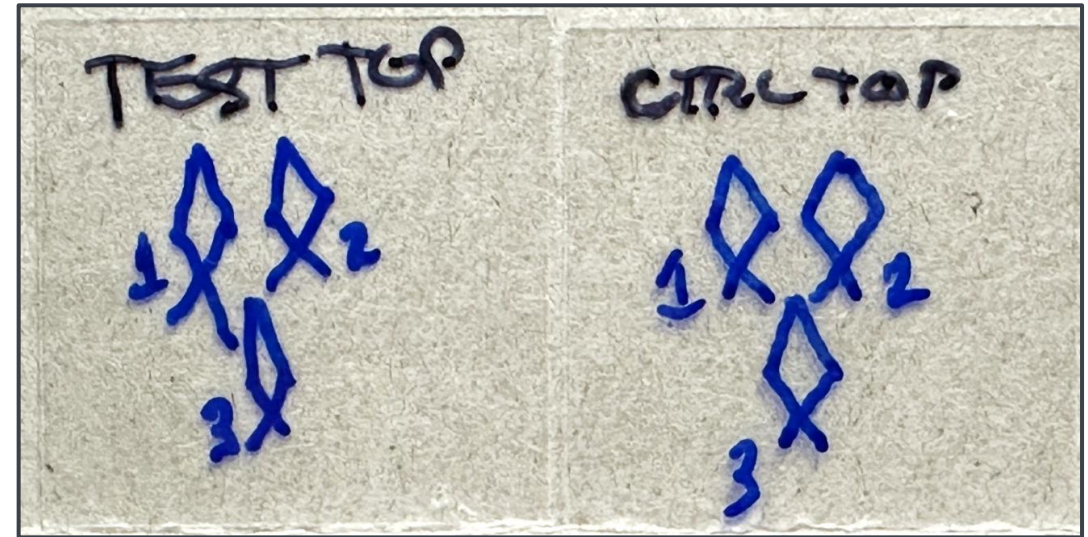
PIPO-SHG can measure pitch angle with sub-degree precision

# PIPO-SHG Details

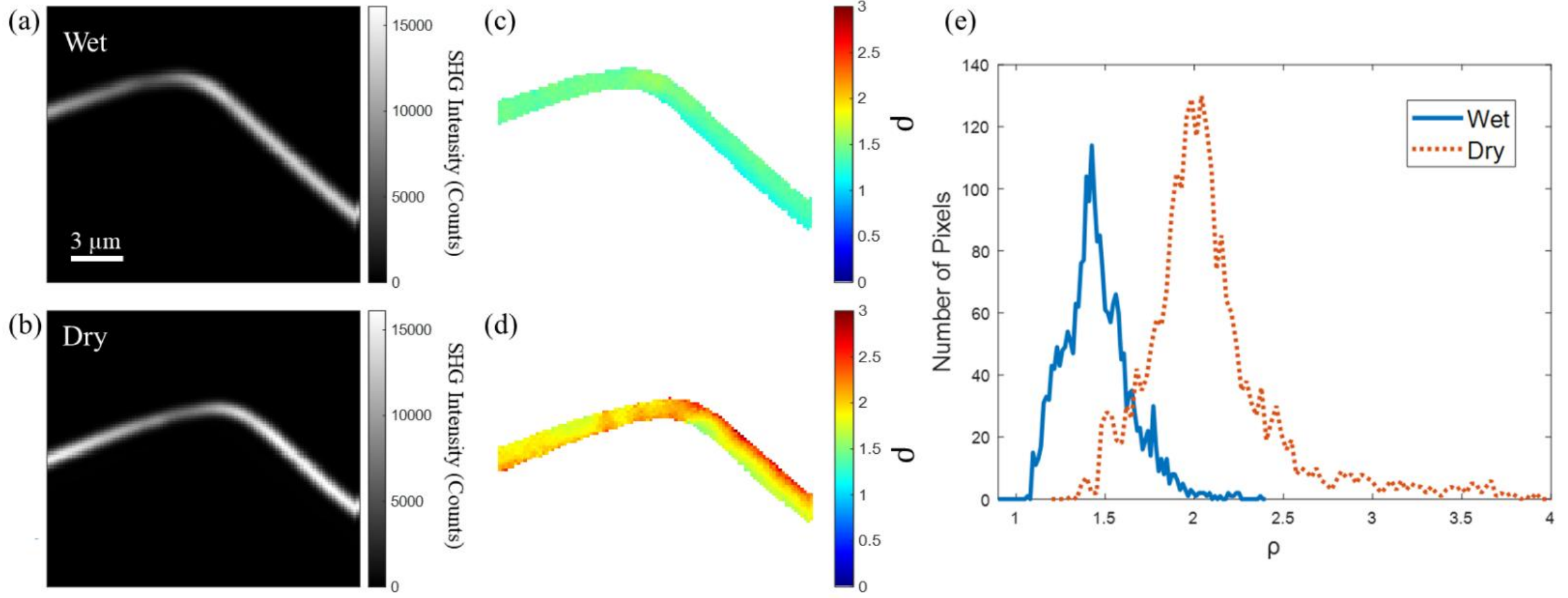
For one experiment:

- 9 TEST images
- 9 CTRL images

- $19 \times 19 \mu\text{m}^2$  image,  $100 \times 100$  pixels
- 25 frames per image in the stack
- Laser power adjusted to maximize counts
- Fit afterwards to extract  $\rho$  and angle



# Typical Ranges for $\rho$ in Collagen Fibrils



Harvey et. al., Nanophotonics, (2023)